

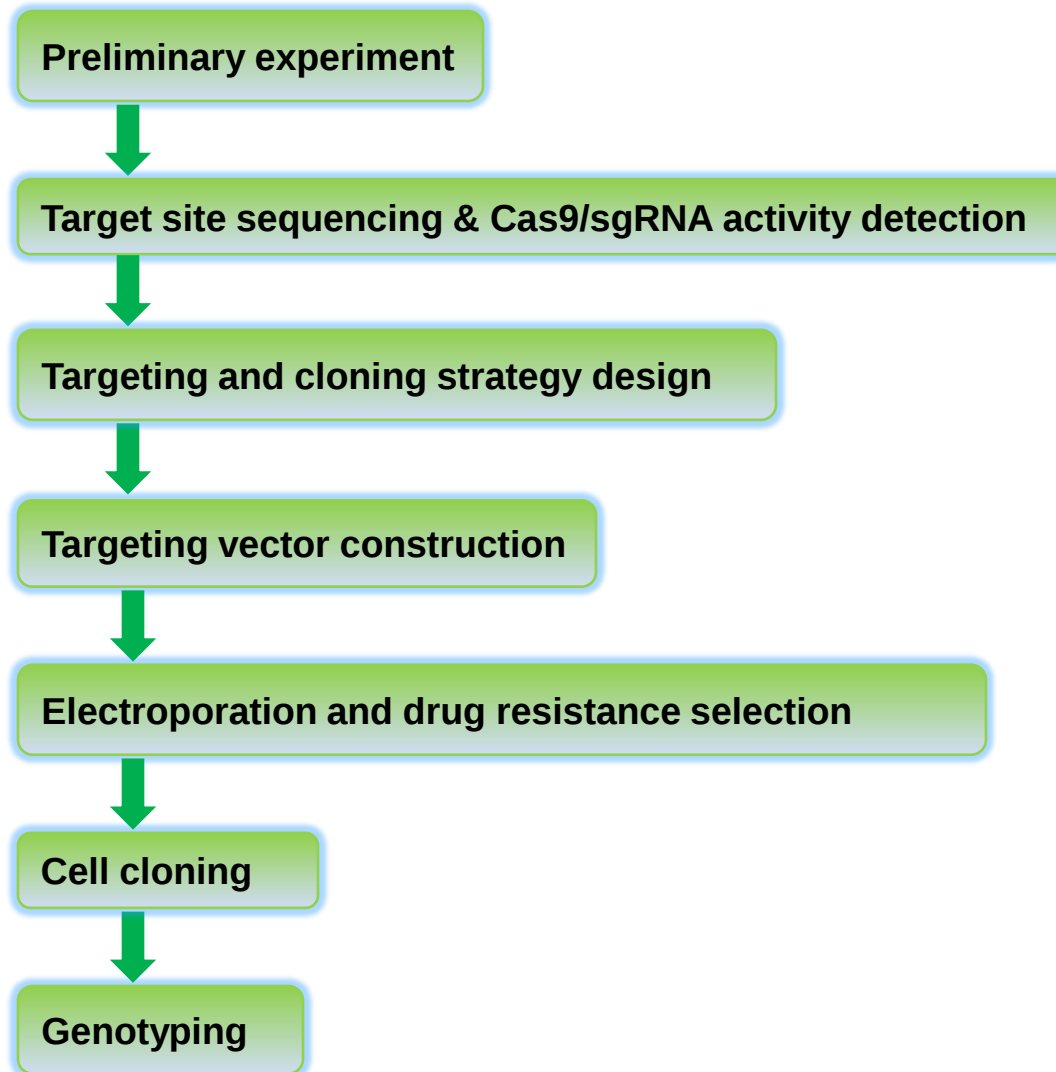


Project status report

2017.05.25~2018.08.15

Project: CL-CYH-008 KI S18 cell line

Work Flow



Part 1

Preliminary experiment

Titration of Antibiotic

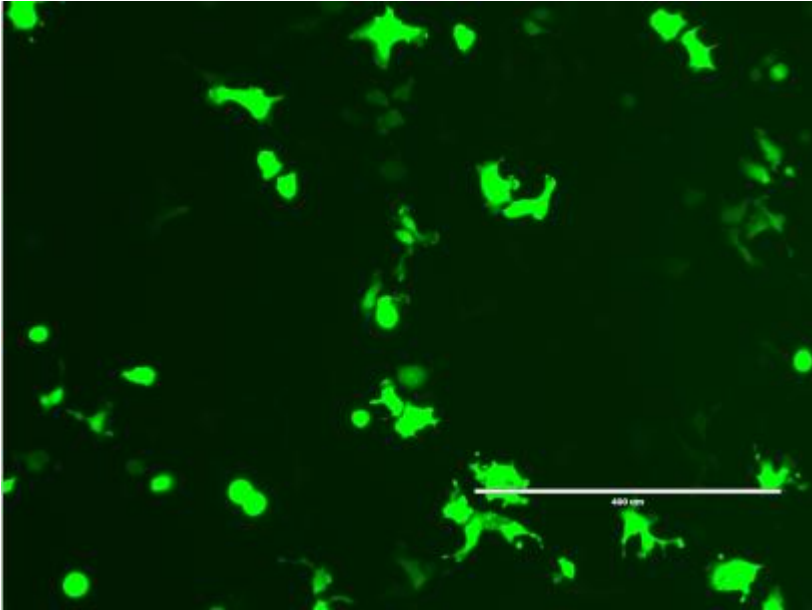
Antibiotic	Concentration ($\mu\text{g/mL}$)
Puromycin	0.5
G418	1200

Transfection Efficiency Optimisation

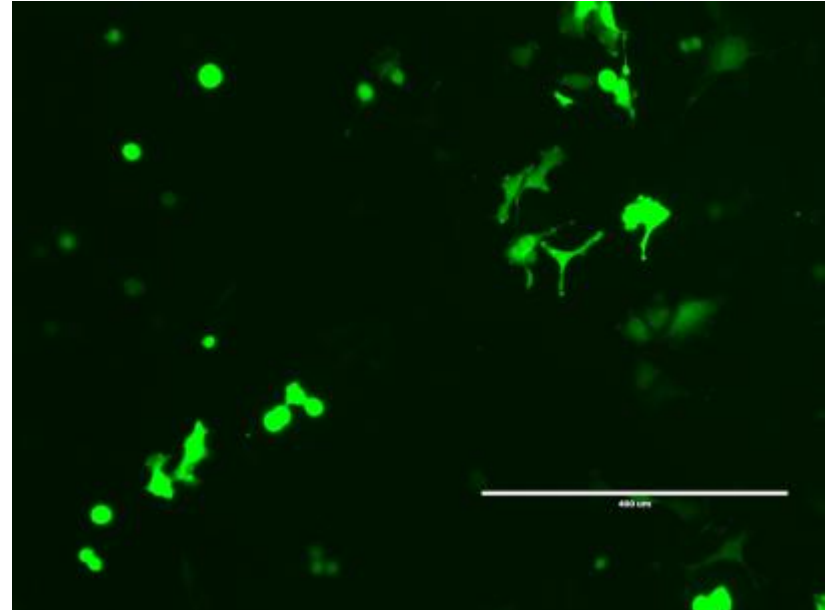
No.	Pulse Voltage	Pulse Width	Pulse no.	Transfection Efficiency	Cell Viability
1	1000	40	1	29.9%	70%
2	900	40	1	23.3%	73%

FACS analysis

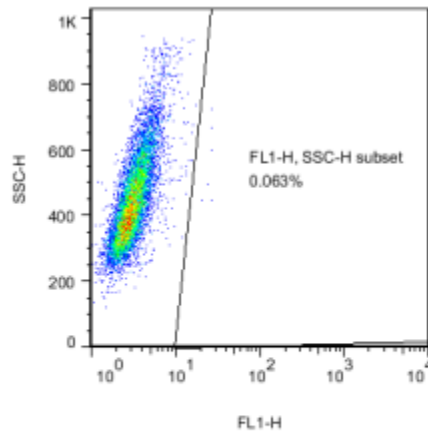
900v 40ms 1pulse



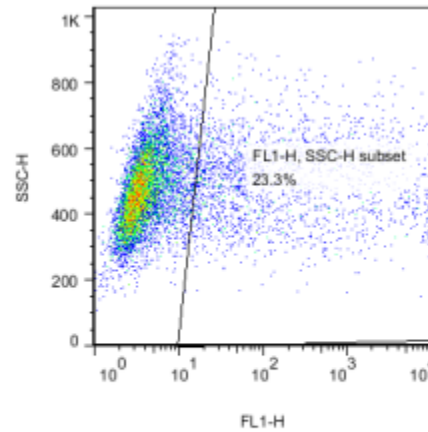
1000v 40ms 1pulse



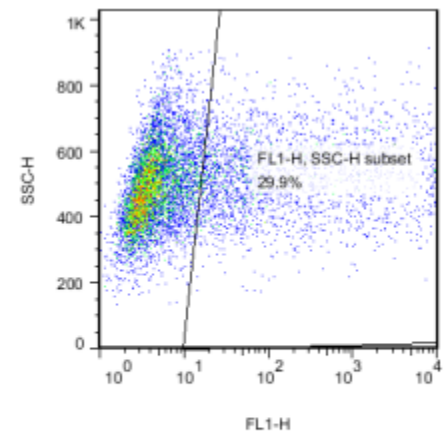
FACS analysis



S18-WT.006
FSC-H, SSC-H subset
9525



S18-900-40-1.008
FSC-H, SSC-H subset
9209



S18-1000-40-1.007
FSC-H, SSC-H subset
9173

Conclusion

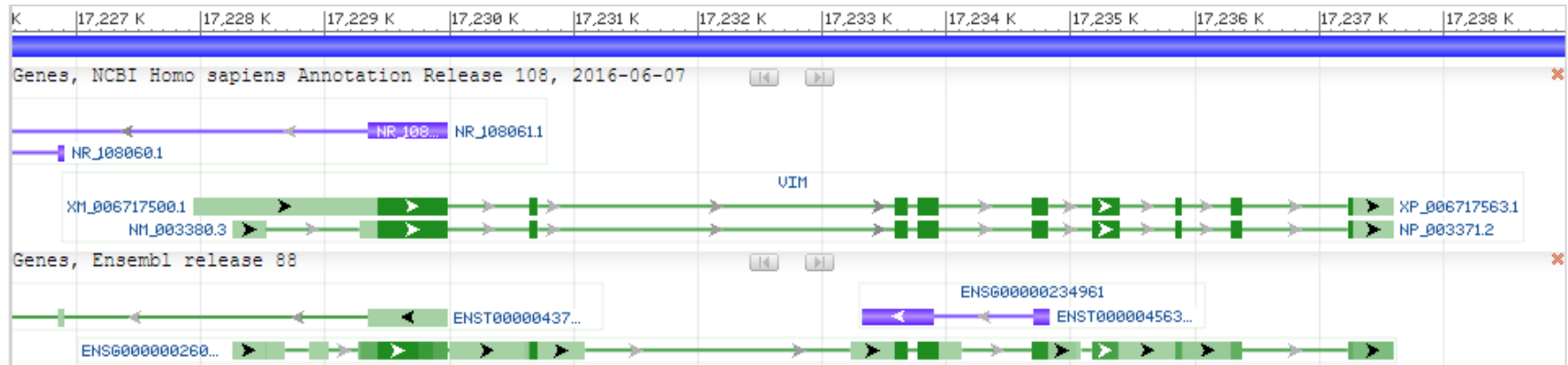
S18 cell line can be modified.

Part 2

Targeting and cloning strategy design

I. Background

1. Gene ID: 7431
2. Human *CL-CYH-008* gene spans about 9.7kb on chromosome 10 forward strand.

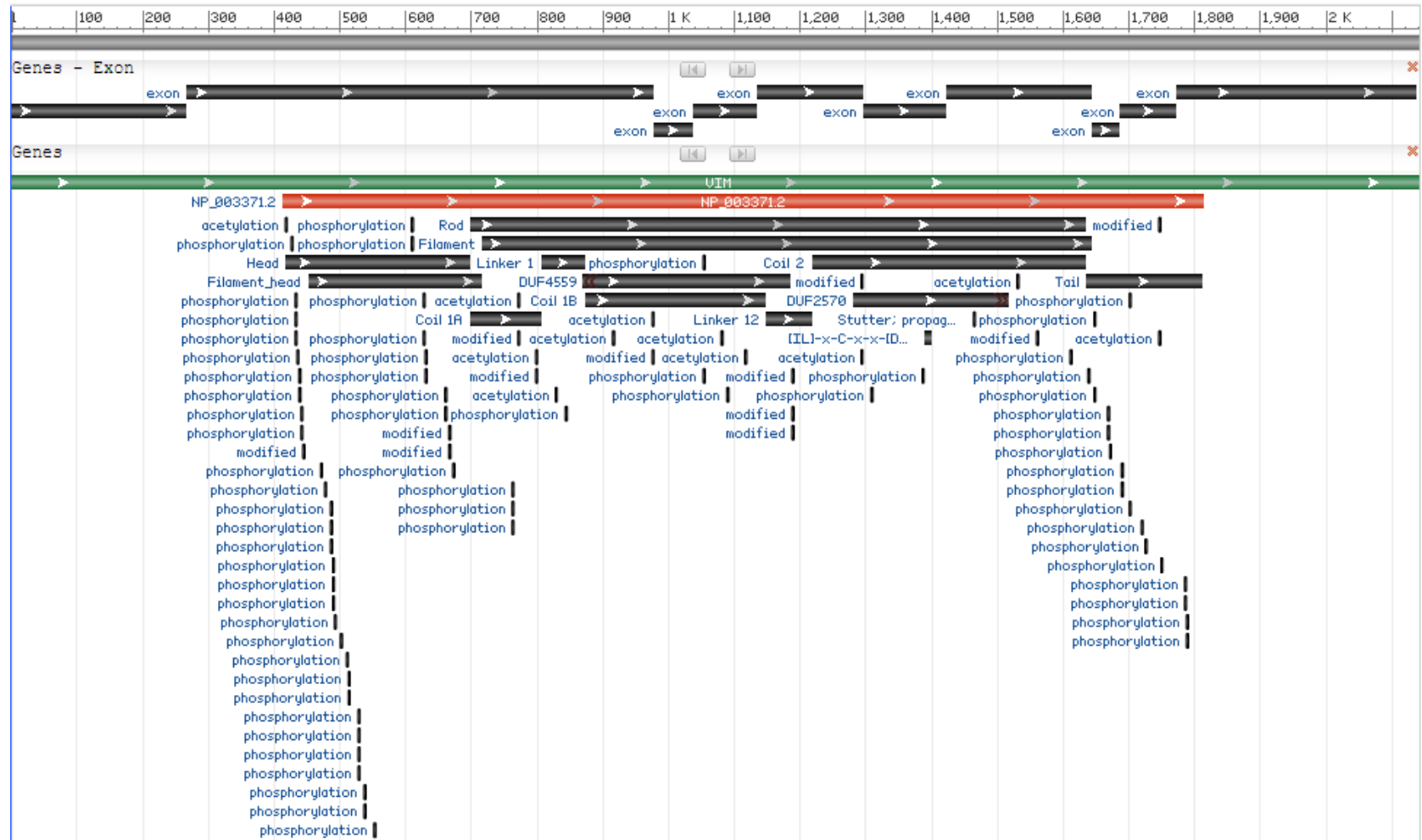


I. Background

3. There are 10 transcripts in this gene.

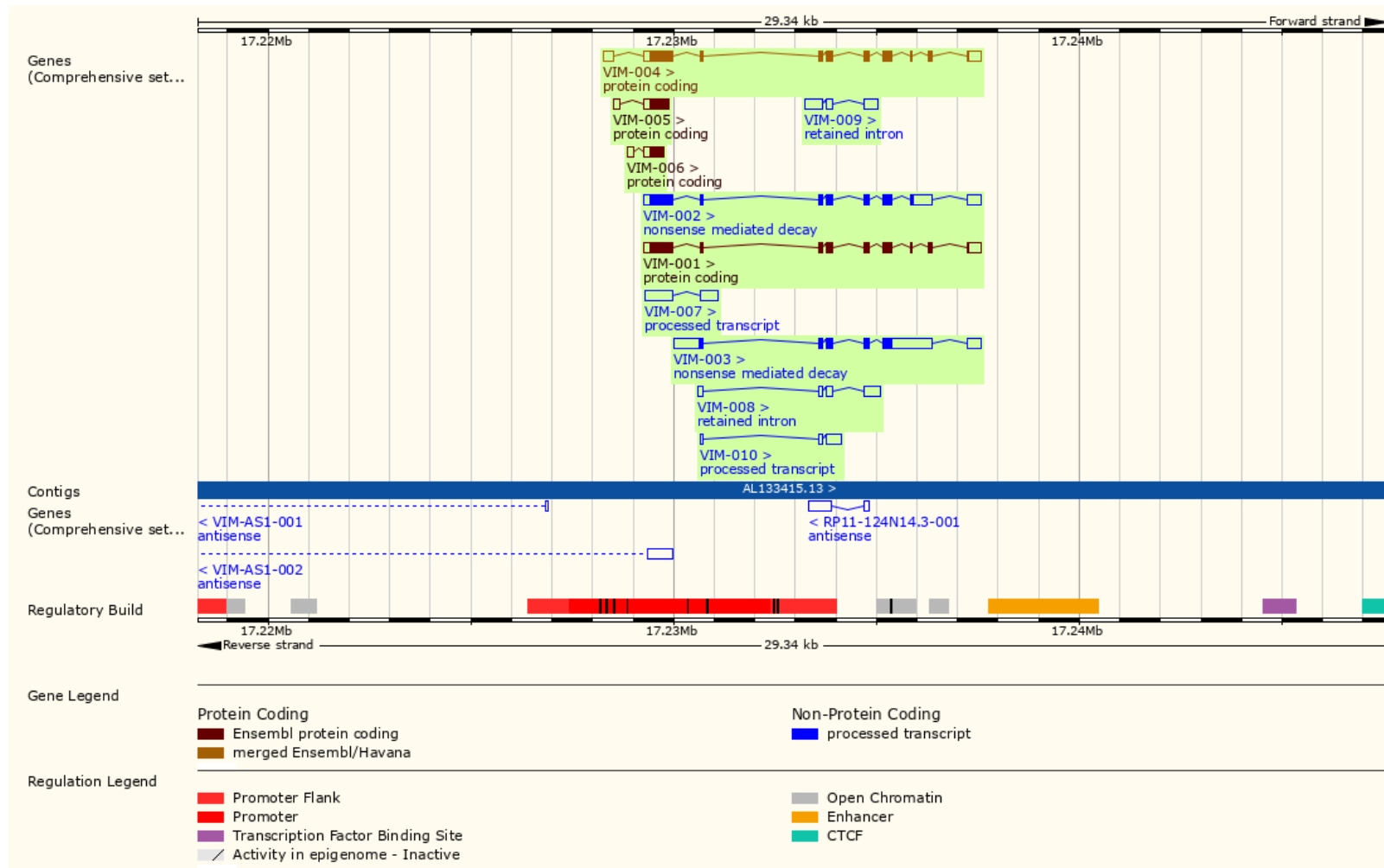
Name	Transcript ID	bp	Protein	Biotype	CCDS	UniProt	RefSeq	Flags
VIM-004	ENST00000544301.6	2135	466aa	Protein coding	CCDS7120	P08670 V9HWE1	NM_003380 NP_003371	TSL:1 APPRIS P1
VIM-001	ENST00000224237.9	1868	466aa	Protein coding	CCDS7120	P08670 V9HWE1	-	TSL:1 GENCODE basic APPRIS P1
VIM-005	ENST00000478746.1	757	149aa	Protein coding	-	A0A1B0GTT5	-	CDS 3' incomplete TSL:2
VIM-006	ENST00000497849.1	634	109aa	Protein coding	-	A0A1B0GVG8	-	CDS 3' incomplete TSL:2
VIM-003	ENST00000469543.5	2666	228aa	Nonsense mediated decay	-	B0YJC5	-	TSL:2
VIM-002	ENST00000487938.5	2272	431aa	Nonsense mediated decay	-	B0YJC4	-	TSL:5
VIM-007	ENST00000485947.1	1139	No protein	Processed transcript	-	-	-	TSL:2
VIM-010	ENST00000637053.1	508	No protein	Processed transcript	-	-	-	TSL:2
VIM-009	ENST00000495528.1	962	No protein	Retained intron	-	-	-	TSL:2
VIM-008	ENST00000421459.2	753	No protein	Retained intron	-	-	-	TSL:2

4. Domains and features



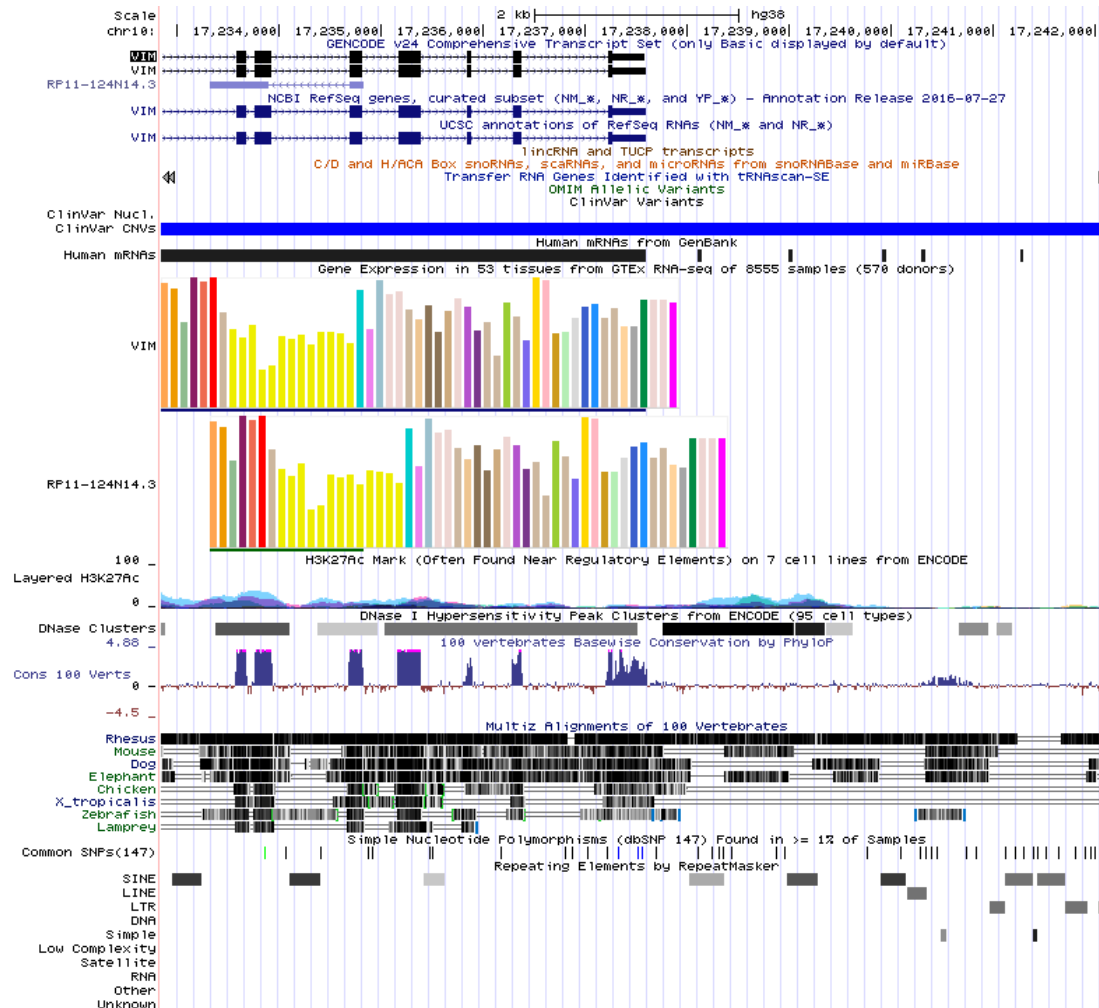
I. Background

5. Regulation



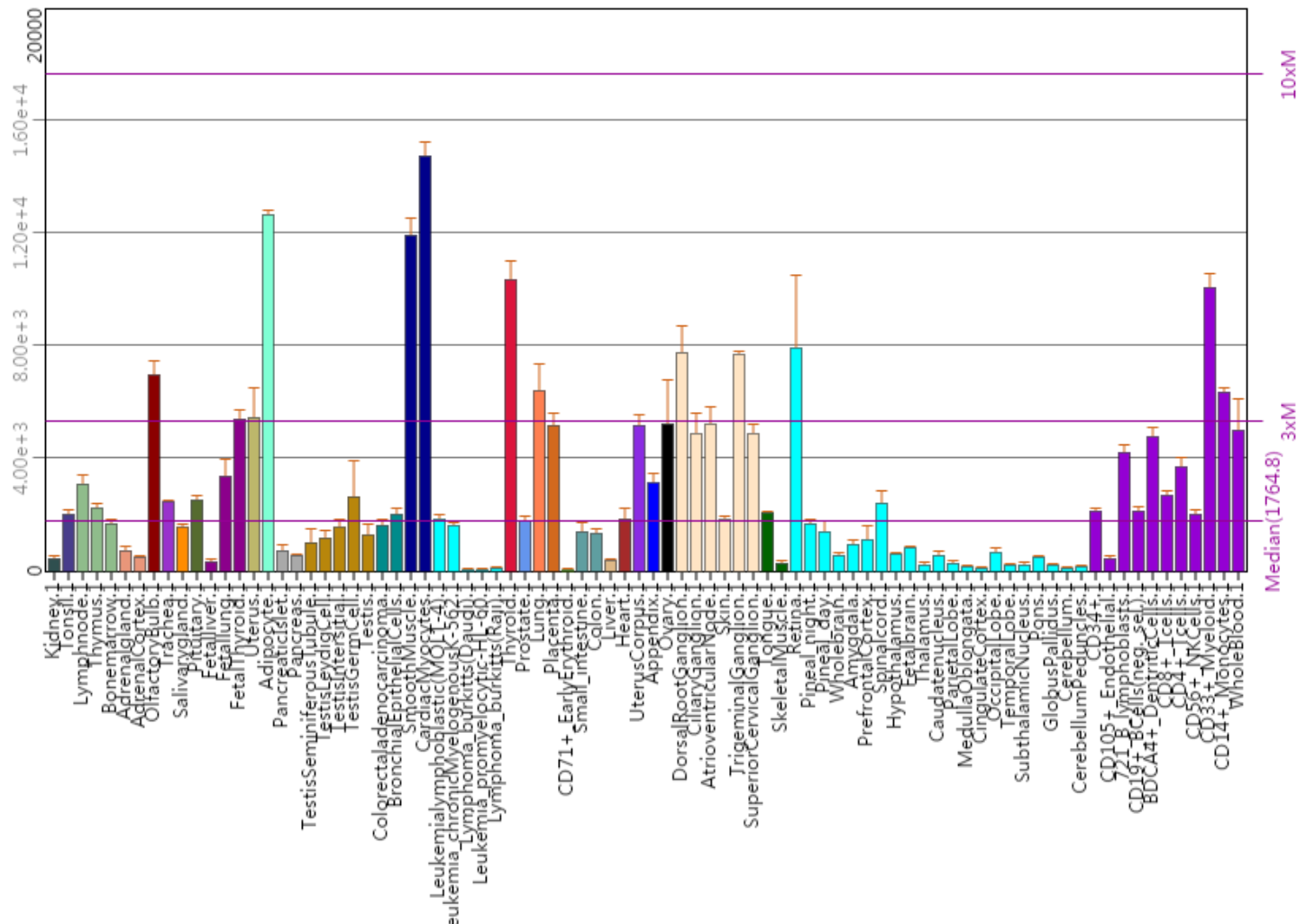
I. Background

6. Conservation



I. Background

7. Expression pattern (from BioGPS)



I. Background

8. Existing mouse models (from MGI)

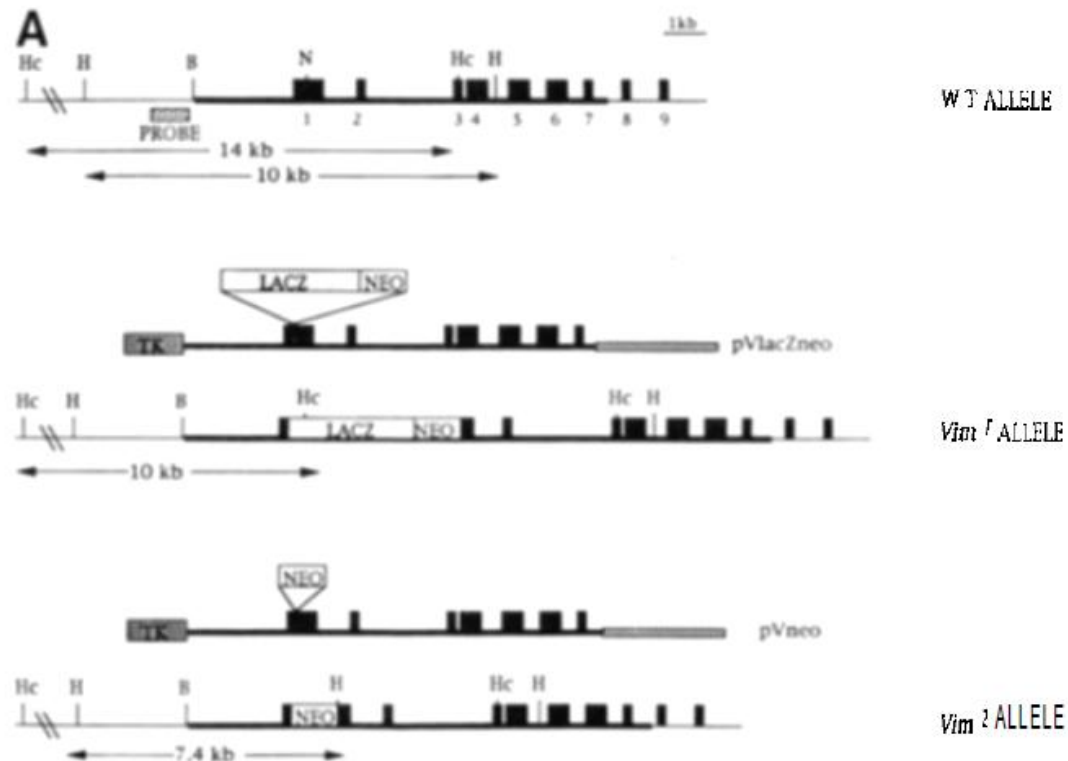
Allele Symbol Gene; Allele Name	Chr	Synonyms	Category	Abnormal Phenotypes Reported in these Systems	Human Disease Models
Vim^{tm1Cba} vimentin; targeted mutation 1, Charles Babinet	2	Vim- Vim1	Targeted (Null/knockout, Reporter)	behavior, cardiovascular, cellular, hematopoietic, homeostasis, immune, mortality/aging, muscle, nervous system	
Vim^{tm2Cba} vimentin; targeted mutation 2, Charles Babinet	2	Vim- Vim2	Targeted (Null/knockout)	cardiovascular, cellular	
Vim^{tm1(KOMP)Vlcg} vimentin; targeted mutation 1, Velocigene	2		Targeted (Null/knockout, Reporter) (Cell Line)		
Vim^{tm1a(EUCOMM)Wtsi} vimentin; targeted mutation 1a, Wellcome Trust Sanger Institute	2		Targeted (Conditional ready, Null/knockout, Reporter) (Cell Line)		
Vim^{tm1e(EUCOMM)Wtsi} vimentin; targeted mutation 1e, Wellcome Trust Sanger Institute	2		Targeted (Null/knockout, Reporter) (Cell Line)		

<http://www.informatics.jax.org/allele/summary?markerId=MGI:98932&alleleType=Targeted>

I. Background

9. Targeting strategy of existing models

CL-CYH-008^{tm1Jjon}

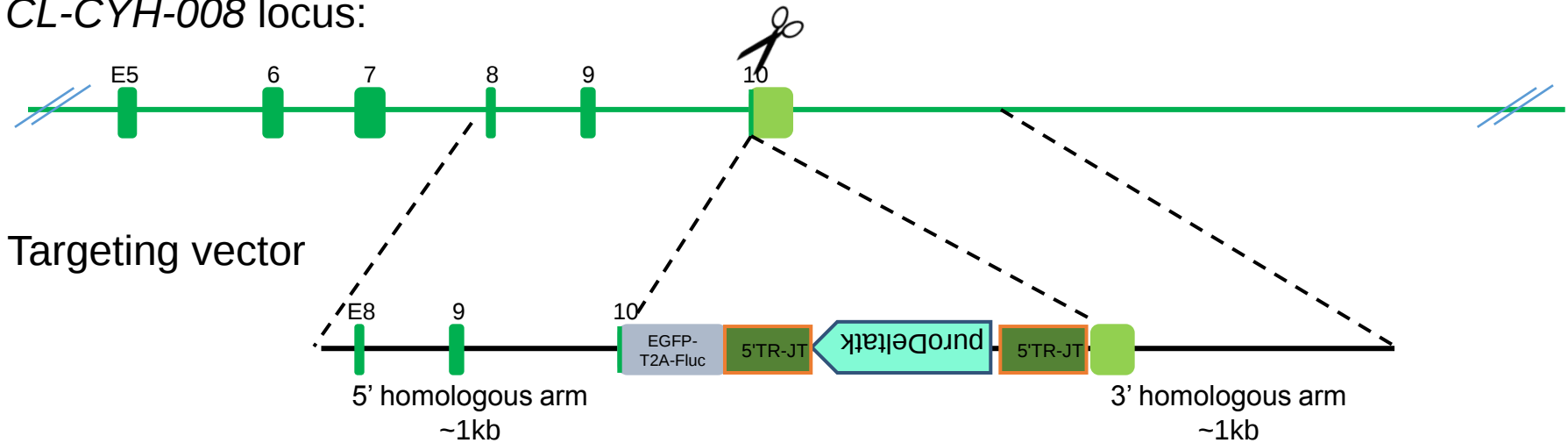


Note:

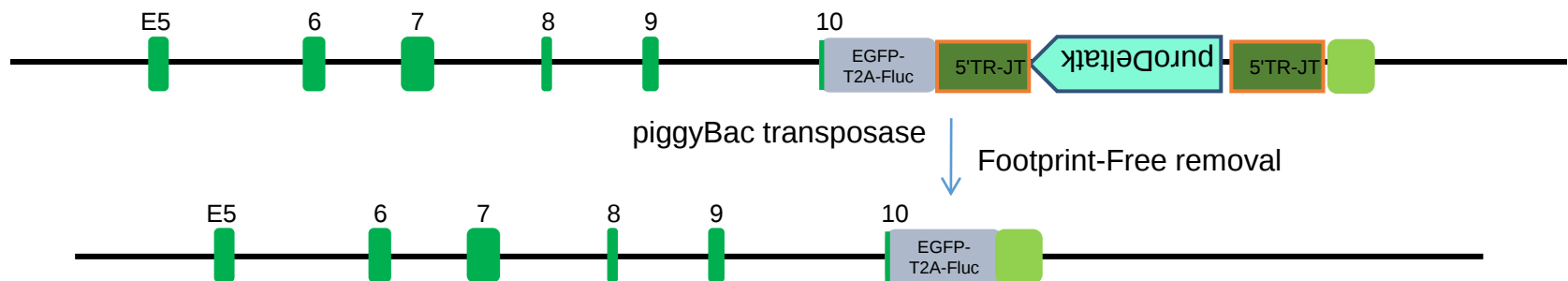
The design for CL-CYH-008^{tm1Jjon} is based on Derksen PW, et al., Somatic inactivation of E-cadherin and p53 in mice leads to metastatic lobular mammary carcinoma through induction of anoikis resistance and angiogenesis. Cancer Cell. 2006 Nov;10(5):437-49.

II. Targeting strategy (1) - *EGE(CRISPR/Cas9)* system

CL-CYH-008 locus:



Targeted allele

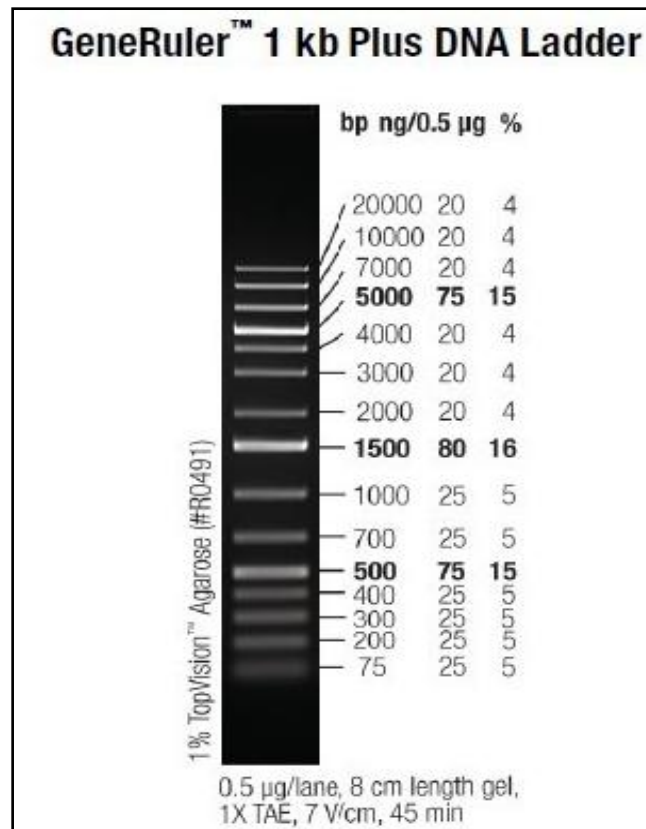


Note: This design is based on transcript 004 ([NM_003380](#)).



Marker used in the experiments

DNA electrophoresis

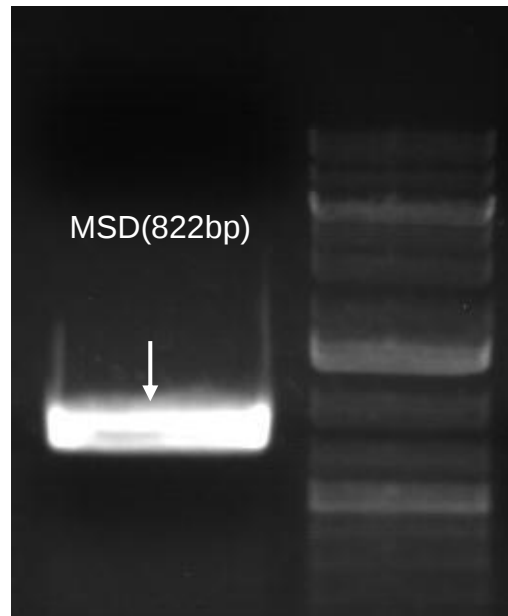


Part 3:
Cas9/sgRNA plasmid construction
&
UCA™ assay

I. Sequencing primer design of the target site

Primer	Sequence (5'-3')	Tm (°C)	Product size (bp)
CL-CYH-008-MSD-F	GCATCAAGCTTGGTACCGATTGCTTTTAACT TGGCTGTATTGTGT	64	822
CL-CYH-008-MSD-R	ACTTAATCGTGGAGGATGATACTACATCTCAA GCATAAATAACCTGA	65	

PCR and sequencing of potential sgRNA target site:



Conclusion: The PCR products (S18 cell line) were sequenced and confirmed as same as those from the NCBI and Ensembl database.

II. *sgRNA design*

Guide	Score	Sequence (5'-3')
Guide #1	76	TCAGGAGCGCAAGATAGATT TGG
Guide #2	76	ACAAGACCCTCTTCCACTAC AGG
Guide #3	76	CAAGTTGGTTGGATACTTGC TGG
Guide #4	73	CAGCAAGTATCCAACCAACT TGG
Guide #5	68	GCGCAAGATAGATTTGGAAT AGG
Guide #6	67	CAGGCAATATAAGGATCCAG TGG
Guide #7	63	CTGCATGAGTGAAAACCTAGA AGG

CRISPR design tool : <http://crispr.mit.edu/>

III. *GuideRNA sequence*

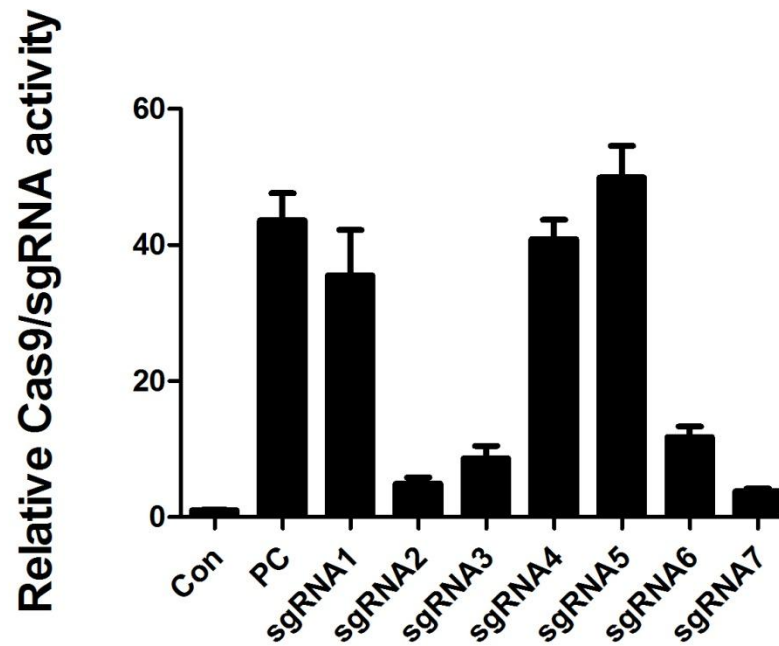
sgRNA	GuideRNA sequence
CL-CYH-008-sgRNA1	GGAGCGCAAGATAGATT
CL-CYH-008-sgRNA2	GGACAAGACCCTCTTCCACTAC
CL-CYH-008-sgRNA3	GGTTGGTTGGATACTTGC
CL-CYH-008-sgRNA4	GGCAAGTATCCAACCAACT
CL-CYH-008-sgRNA5	GGCGCAAGATAGATTTGGAAT
CL-CYH-008-sgRNA6	GGCAATATAAGGATCCAG
CL-CYH-008-sgRNA7	GGCATGAGTGAAACTAGA

Cas9/sgRNA plasmid construction

Cas9/sgRNA plasmid construction was completed and the sequences were further confirmed by DNA sequencing.

UCATM assay:

Note: UCA (Universal CRISPR Activity Assay), a sgRNA activity detection system developed by Biocytogen, is simpler and more sensitive than MSDase assay.



Conclusion: the sgRNA5 was used for next step.

Part 4:

Targeting vector construction

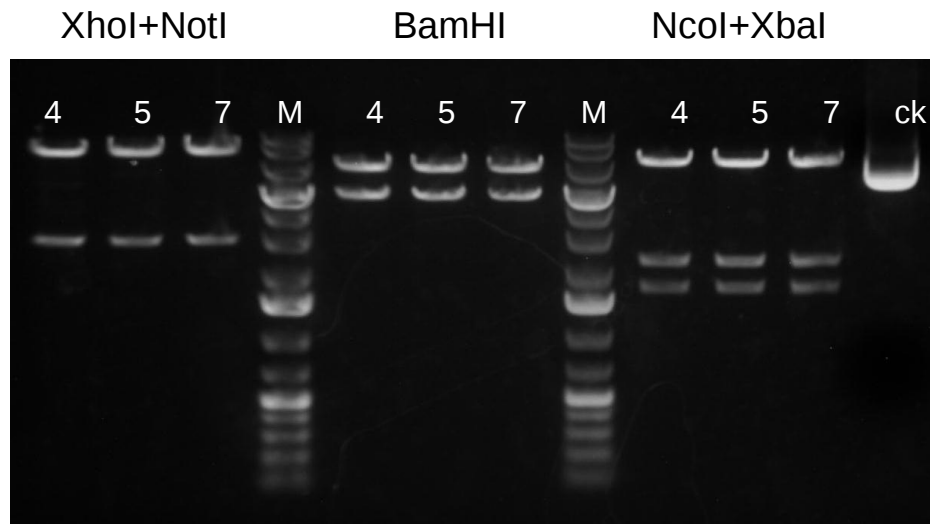
I. *Cloning primer design*

Primer	Sequence (5'-3')	Restriction enzyme	Tm(°C)	Product size(bp)	Template
CL-CYH-008-LR-F	atcgCTCGAGTTGGCGAGTAGTACTTTACACAATT	XhoI	65	8164	Synthesis
CL-CYH-008-RR-R	cagtGCGGCCGC TAGCTATCACACACACACAATAA	NotI	69		

Fragment LR-A-RR (Synthesis) will be ligated into LScKO-4G vector

Restriction analysis: CL-CYH-008 KI targeting vector

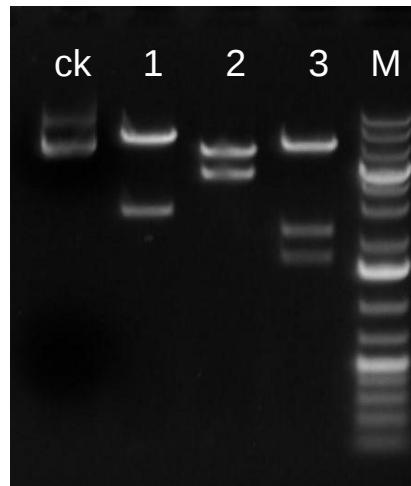
1. Restriction enzyme: XhoI+NotI
Expected products: 8149bp+2773bp
2. Restriction enzyme: BamHI
Expected products: 6419bp+4503bp
3. Restriction enzyme: NcoI+XbaI
Expected products: 7118bp+2198bp+1606bp



#4, #5 and #7 clones were correct. #5 and #7 were sent for DNA sequencing. Sequences of #5 and #7 were correct.

Restriction analysis: CL-CYH-008 KI targeting vector(#7) after maxi-prep

1. Restriction enzyme: XhoI+NotI
Expected products: 8149bp+2773bp
2. Restriction enzyme: BamHI
Expected products: 6419bp+4503bp
3. Restriction enzyme: NcoI+XbaI
Expected products: 7118bp+2198bp+1606bp



Note: The number labeled in the pictures indicates the group number of restriction enzyme showed above.

Part 5

Electroporation & Cell cloning

Electroporation and clone screening

- sgRNA5 and CL-CYH-008-targeting vector were electroporated into S18 cell line.
- Drug resistance selection.
- Screening of mix colonies by PCR.
- Clonal growth of cells in semisolid media.
- 96 resistant clones were picked and expanded for further analysis.
- 96 colonies were ready for genotyping.

Electroporation and clone screening

- CL-CYH-008-piggyBac vector was electroporated into mix colonies.
- Drug resistance selection.
- Screening of mix colonies by PCR.
- Clonal growth of cells in semisolid media.
- 96 resistant clones were picked and expanded for further analysis.
- 96 colonies were ready for genotyping.

Electroporation and clone screening

- PCR screening was performed.
- Amplification and cryopreservation of positive colonies.

Electroporation and clone screening(the 2nd)

- sgRNA5 and CL-CYH-008-targeting vector were electroporated into S18 cell line.
- Drug resistance selection.
- Screening of mix colonies by PCR.
- Clonal growth of cells in semisolid media.
- CL-CYH-008-piggyBac vector was electroporated into mix colonies.
- Drug resistance selection.

Electroporation and clone screening(the 2nd)

- 96 resistant clones were picked and expanded for further analysis.(CL-CYH-008 2#)
- 5 colonies were ready for genotyping. (CL-CYH-008 2#)
- 144 resistant clones were picked and expanded for further analysis. (CL-CYH-008-PB 2#)

Electroporation and clone screening

- 2 colonies were ready for genotyping. (CL-CYH-008 2#)
- PCR screening was performed. (CL-CYH-008 2#)
- Amplification and cryopreservation of positive colonies. (CL-CYH-008 2#)
- 19 colonies were ready for genotyping. (CL-CYH-008-PB 2#)

Electroporation and clone screening

- PCR screening was performed. (CL-CYH-008-PB 2#)
- Amplification and cryopreservation of positive colonies. (CL-CYH-008-PB 2#)
- Clonal growth of cells in semisolid media. (CL-CYH-008 2#)
- 96 resistant clones were picked and expanded for further analysis. (CL-CYH-008 2#)

Electroporation and clone screening

- 95 colonies were ready for genotyping.(CL-CYH-008 2#)
- PCR screening was performed. (CL-CYH-008 2#)
- Amplification and cryopreservation of positive colonies.(CL-CYH-008 2#)
- CL-CYH-008-piggyBac vector was electroporated into 3-B5 . (CL-CYH-008-PB 3#)
- Drug resistance selection. (CL-CYH-008-PB 3#)

Electroporation and clone screening

- Drug resistance selection. (CL-CYH-008-PB 3#)
- Screening of mix colonies by PCR. (CL-CYH-008-PB 3#)
- Clonal growth of cells in semisolid media. (CL-CYH-008-PB 3#)

Electroporation and clone screening

- 96 resistant clones were picked and expanded for further analysis. (CL-CYH-008-PB 3#)
- 26 colonies were ready for genotyping. (CL-CYH-008-PBc 3#)
- PCR screening was performed. (CL-CYH-008-PB 3#)
- CL-CYH-008-piggyBac vector was electroporated into 1-F6. (CL-CYH-008-PB 4#)
- Drug resistance selection. (CL-CYH-008-PB 4#)

Electroporation and clone screening

- Amplification and cryopreservation of positive colonies.(CL-CYH-008-PB 3#)
- Screening of mix colonies by PCR.(CL-CYH-008-PB 4#)
- Clonal growth of cells in semisolid media. (CL-CYH-008-PB 4#)

Electroporation and clone screening

- 96 resistant clones were picked and expanded for further analysis.(CL-CYH-008-PB 4#)
- 96 colonies were ready for genotyping.(CL-CYH-008-PB 4#)

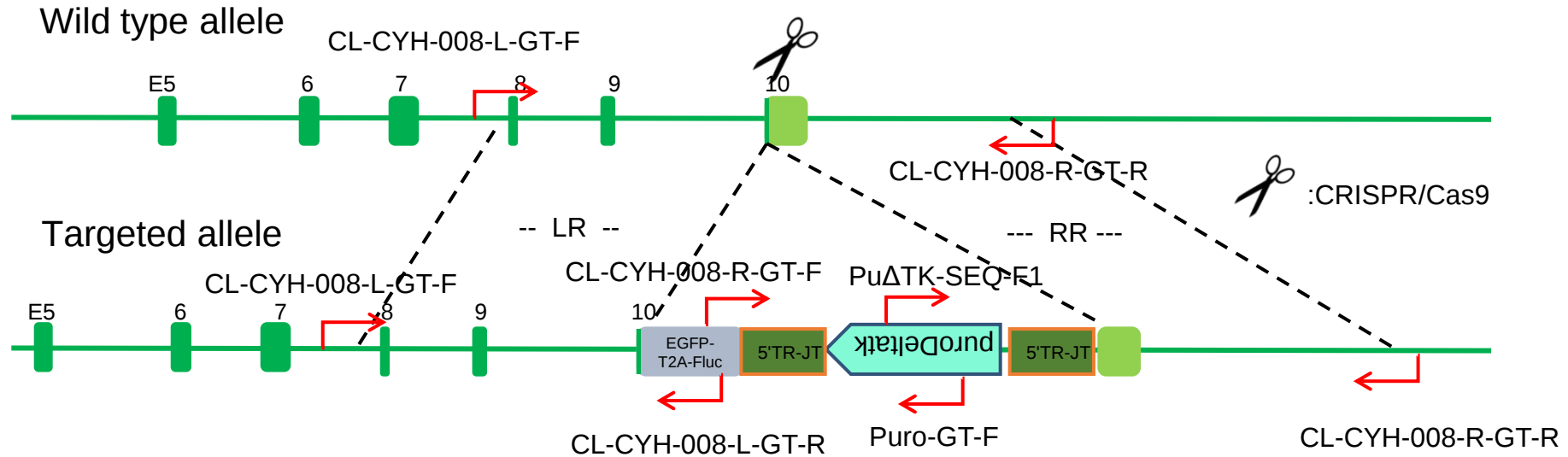
Electroporation and clone screening

- PCR screening was performed.(CL-CYH-008-PB 4#)

Electroporation and clone screening

- Amplification and cryopreservation of positive colonies.(CL-CYH-008-PB 4#)

Screening of mix colonies-Primer design



Screening of mix colonies-Primer design

Primer	Sequence (5'-3')	T _m (°C)	Product size
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut: 2901bp
CL-CYH-008-L-GT-R	GTTGCTTAGGTCGTACTTGTCGATG	59	
CL-CYH-008-R-GT-F	ATGGATAGCAAGACCGACTACCAGG	61	Mut: 3301bp
Puro-GT-F	GCAACAGATGGAAGGCCTCCTGGCG	67	
PuΔTK-SEQ-F1	AGTAGCGTGGGCATGGATCC	61	Mut: 3113bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	59	

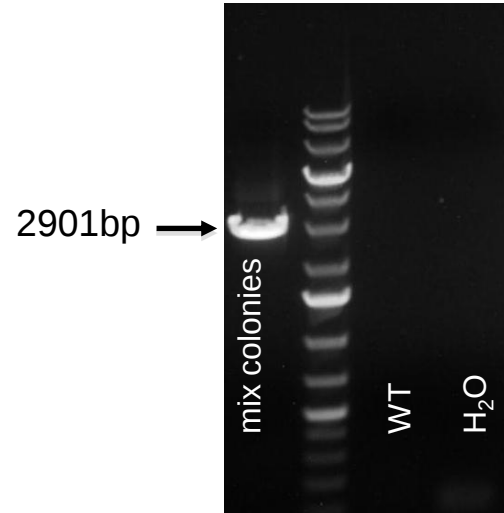
Enzyme: KOD-FX

Program: Touchdown PCR

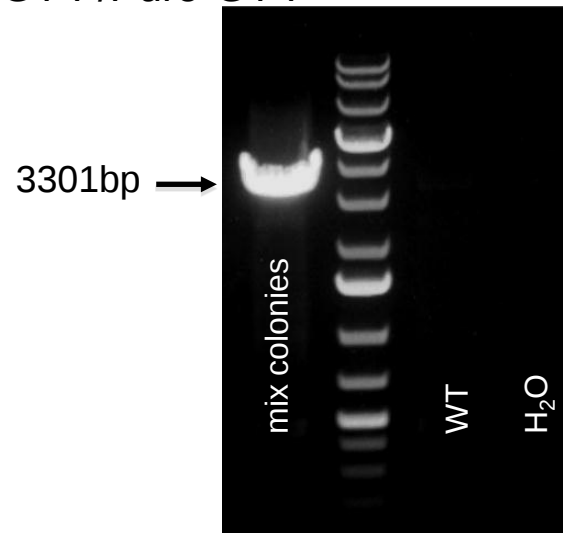
94 °C	2 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

Screening of mix colonies-Genotyping

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

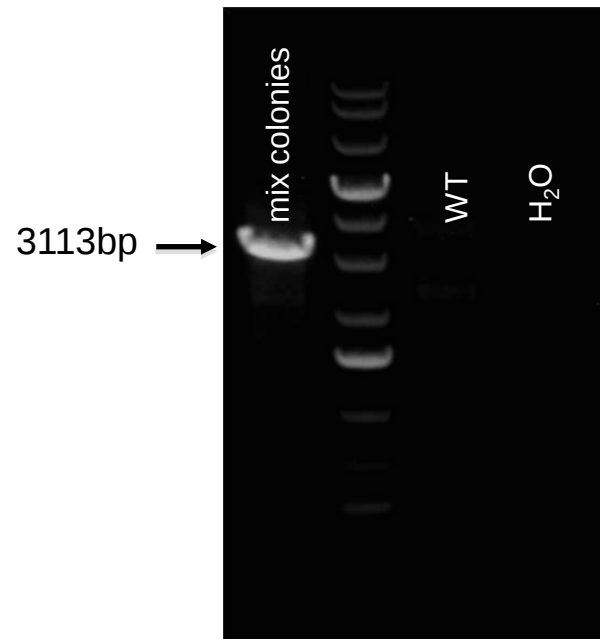


Primers: CL-CYH-008-R-GT-F/Puro-GT-F



Screening of mix colonies-Genotyping

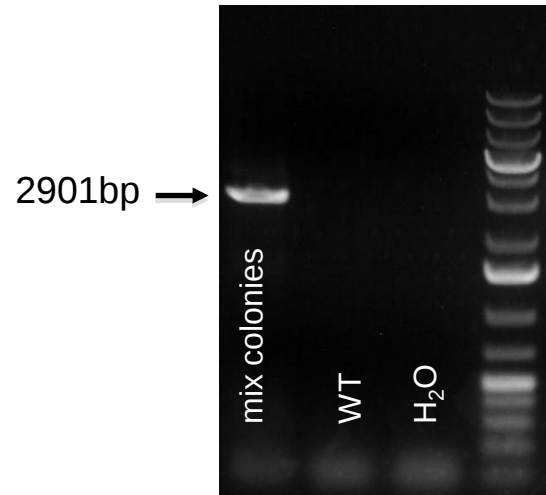
Primers: Pu Δ TK-SEQ-F1/CL-CYH-008-R-GT-R



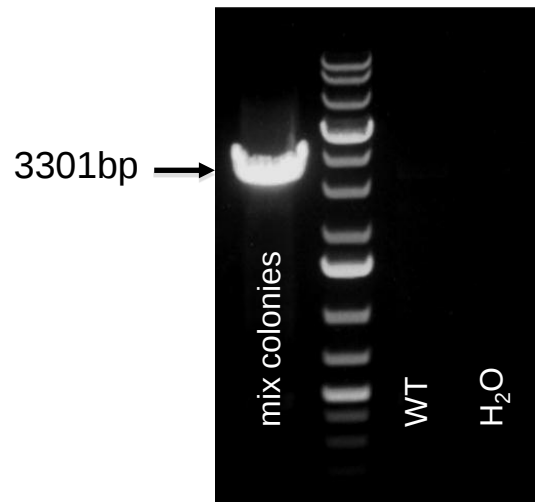
Positive colonies will be picked and expanded for further analysis.

Screening of mix colonies-Genotyping(the 2nd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

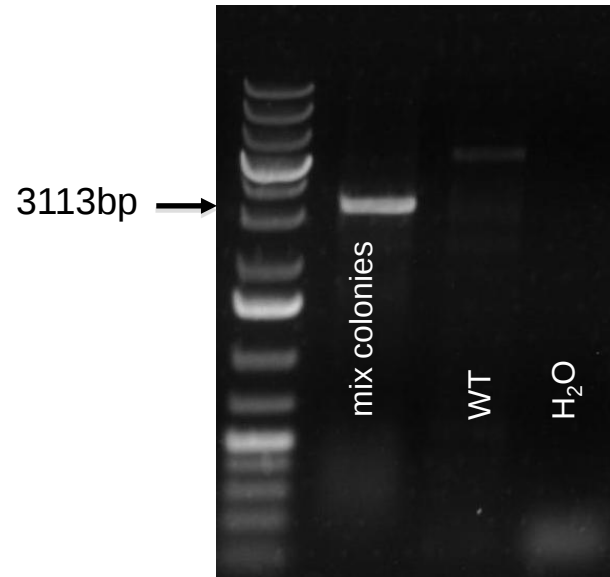


Primers: CL-CYH-008-R-GT-F/Puro-GT-F



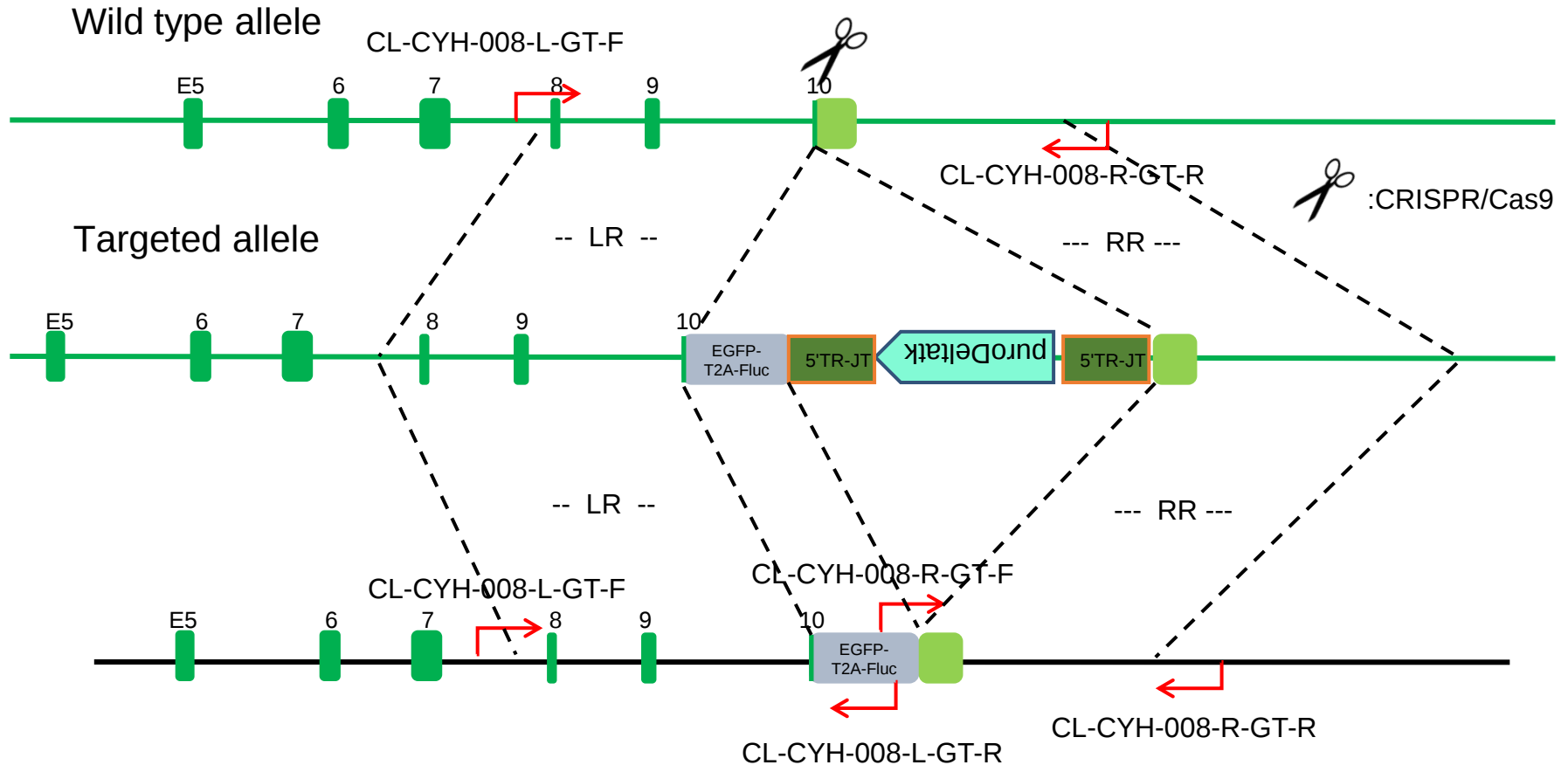
Screening of mix colonies-Genotyping(the 2nd time)

Primers: Pu Δ TK-SEQ-F1/CL-CYH-008-R-GT-R



Positive colonies will be picked and expanded for further analysis.

Screening of mix colonies-piggyBac-Primer design



Screening of mix colonies-piggyBac-Primer design

Primer	Sequence (5'-3')	Tm(°C)	Product size
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut: 2901bp
CL-CYH-008-L-GT-R	GTTGCTTAGGTCGTACTTGTCGATG	59	
CL-CYH-008-R-GT-F	ATGGATAGCAAGACCGACTACCAGG	61	Mut: 2680bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	60	

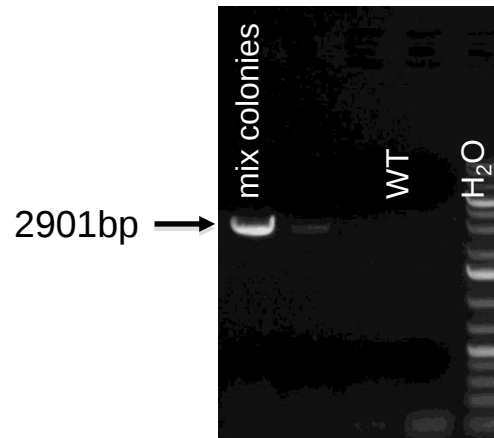
Enzyme: KOD-FX

Program: Touchdown PCR

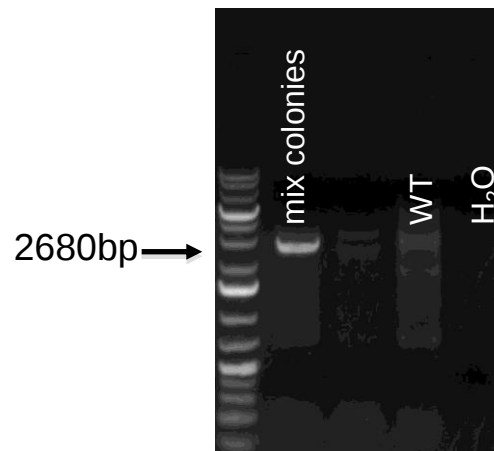
94 °C	2 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57°C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

Screening of *mix colonies*-Genotyping-piggyBac

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R



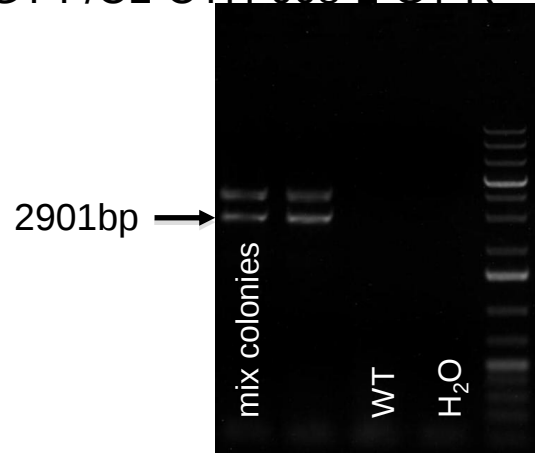
Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R



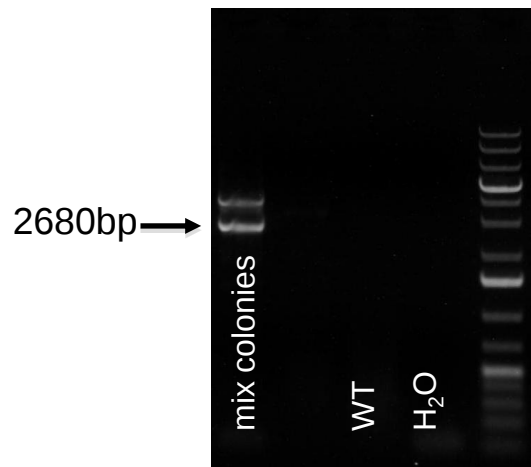
Positive colonies will be picked and expanded for further analysis.

Screening of mix colonies-Genotyping-piggyBac(the 2nd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R



Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

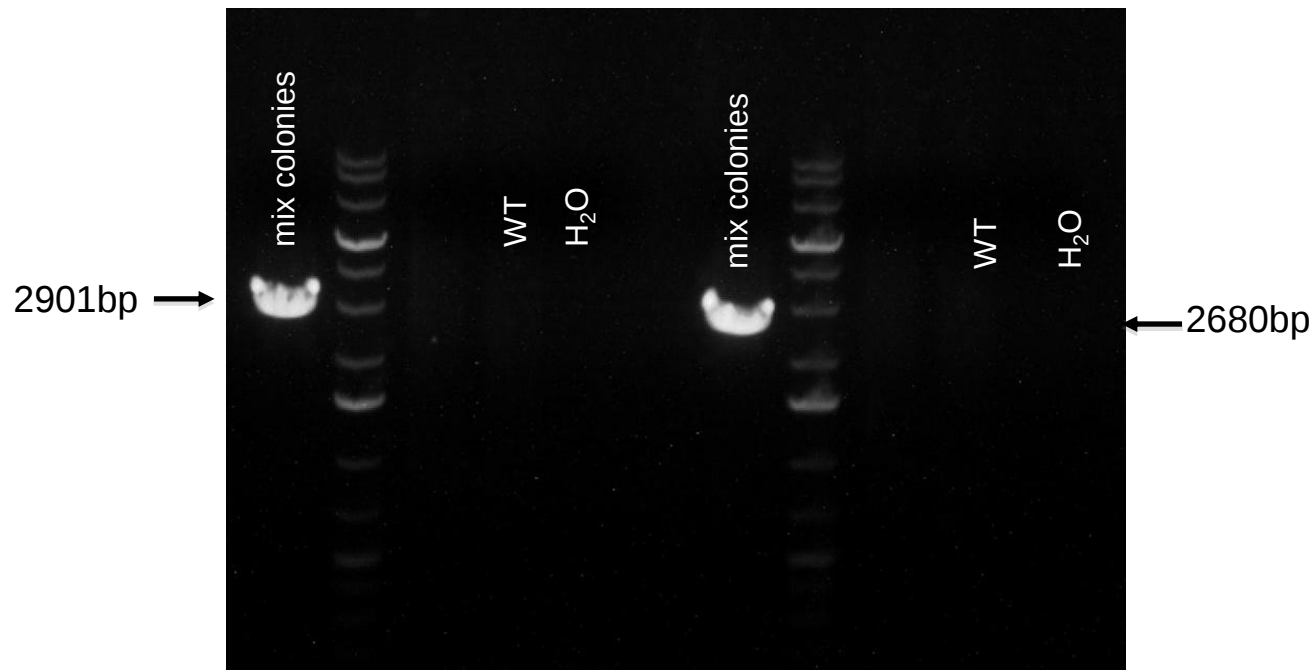


Positive colonies will be picked and expanded for further analysis.

Screening of mix colonies-Genotyping-piggyBac(the 3rd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

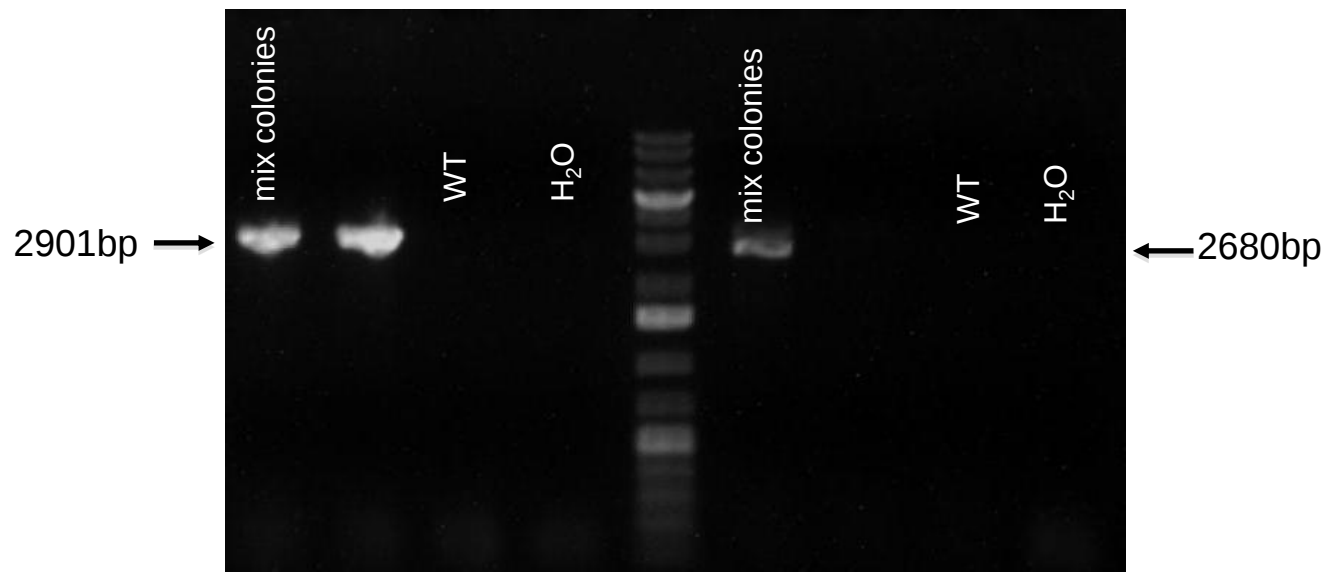


Positive colonies will be picked and expanded for further analysis.

Screening of mix colonies-Genotyping-piggyBac(the 4th time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

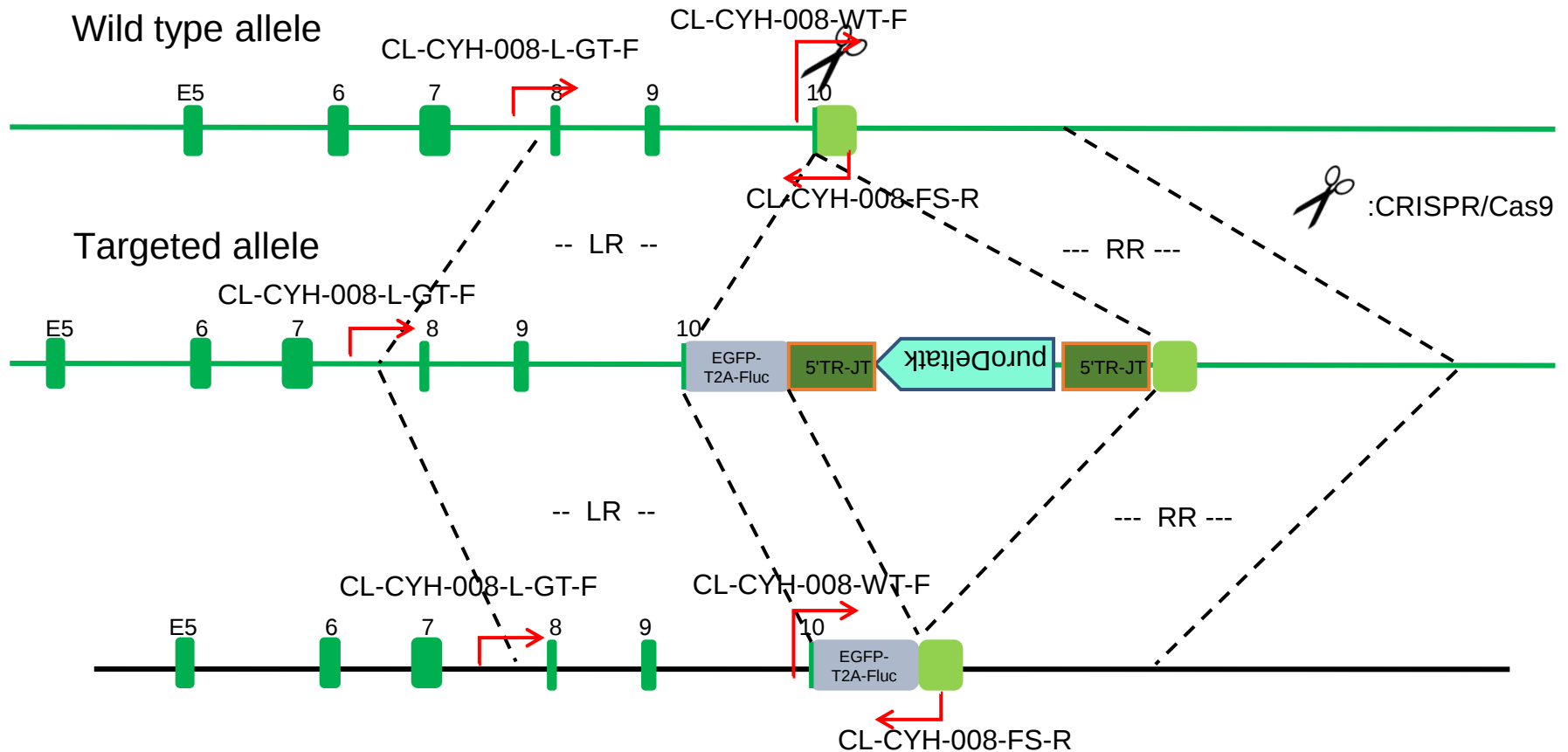


Positive colonies will be picked and expanded for further analysis.

Part 6

Screening of positive colonies

I. Genotyping primer design



I. *Genotyping primer design*

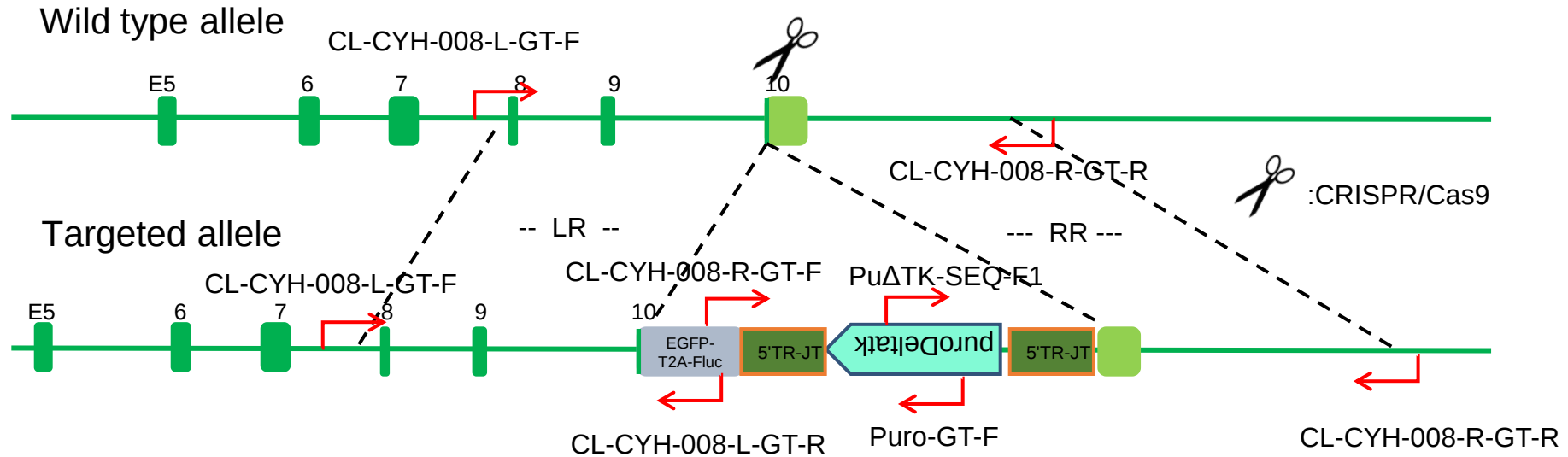
Primer	Sequence (5'-3')	Tm(°C)	Product size
CL-CYH-008-WT-F	GTCTTGTGTGTCTGCCCCTTCTAGT	62	Mut:6406bp WT:527bp Indel:~527bp
CL-CYH-008-FS-R	CCAAAGATTTATTGAAGCAGAACCAAGT	59	
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut:7378bp WT:1499bp Indel:~1499bp
CL-CYH-008-FS-R	CCAAAGATTTATTGAAGCAGAACCAAGT	59	

Enzyme: KOD-FX

Program: Touchdown PCR

94 °C	5 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

II. Genotyping primer design



II. Genotyping primer design

Primer	Sequence (5'-3')	T _m (°C)	Product size
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut: 2901bp
CL-CYH-008-L-GT-R	GTTGCTTAGGTCGTACTTGTCGATG	59	
CL-CYH-008-R-GT-F	ATGGATAGCAAGACCGACTACCAGG	61	Mut: 3301bp
Puro-GT-F	GCAACAGATGGAAGGCCTCCTGGCG	67	
PuΔTK-SEQ-F1	AGTAGCGTGGGCATGGATCC	61	Mut: 3113bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	59	

Enzyme: KOD-FX

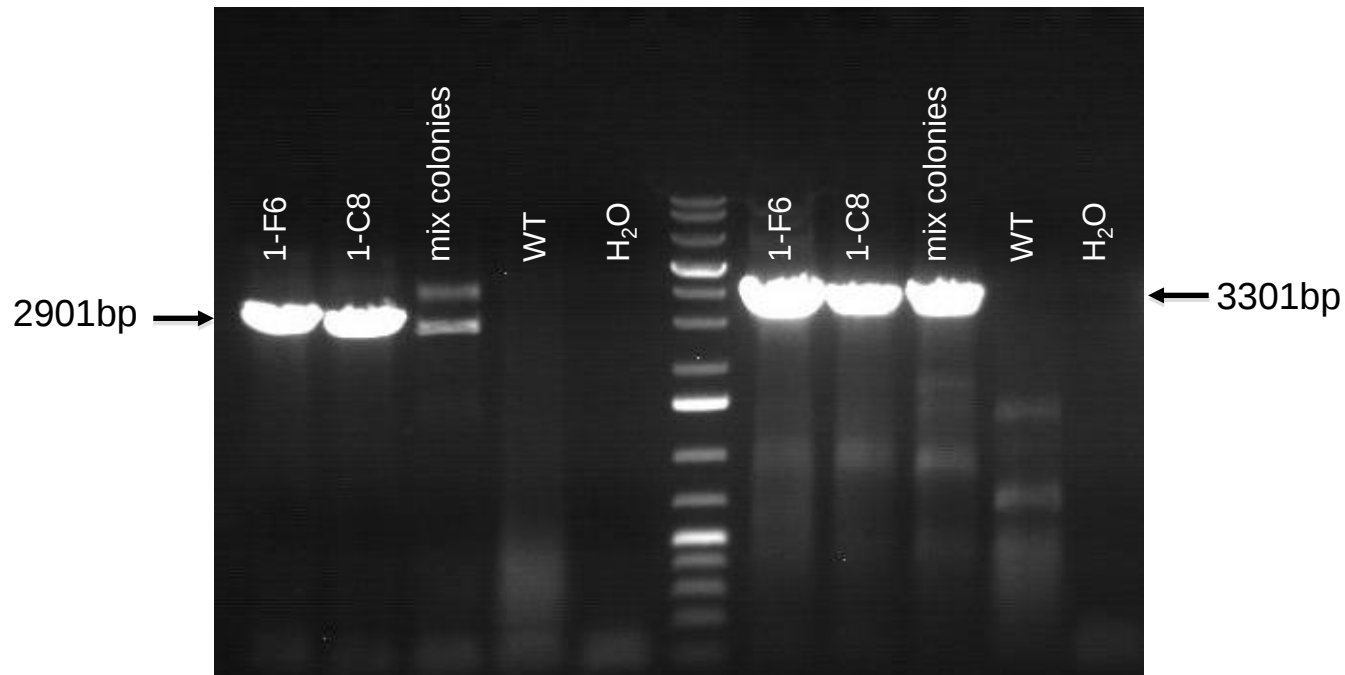
Program: Touchdown PCR

94 °C	5 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

Genotyping-Junction PCR

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

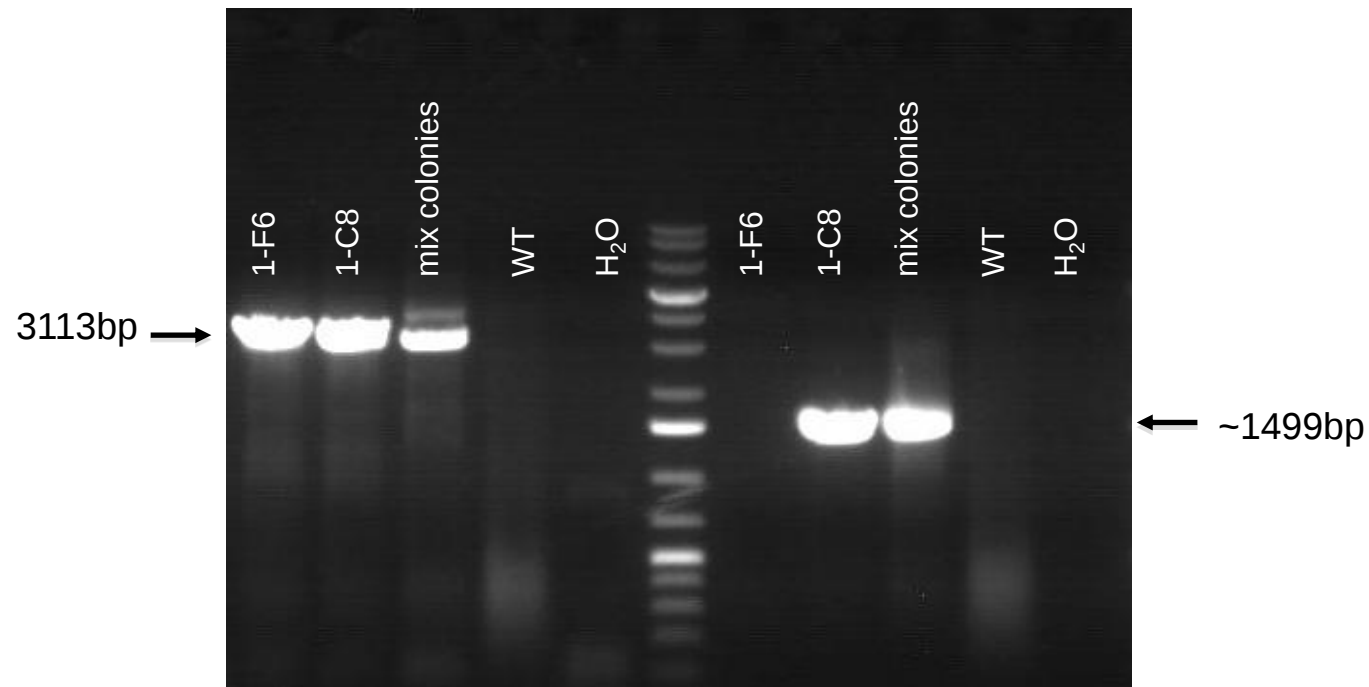
Primers: CL-CYH-008-R-GT-F/Puro-GT-F



Genotyping-Junction PCR

Primers: Pu Δ TK-SEQ-F1/CL-CYH-008-R-GT-R

Primers: CL-CYH-008-L-GT-F/ CL-CYH-008-FS-R



Colony screening by PCR and sequencing

Abbreviation:

“no band”: No expected band was detected in PCR screening, but integration band was detected in the same reaction.

HR: Homologous Recombination.

Δ: deletion.

Colonies screening by PCR and sequencing

Colony	HR allele	non-HR allele	Genotype
1-F6	+	none	KI,KI
1-C8	+	Δ 6bp	KI, Δ 6

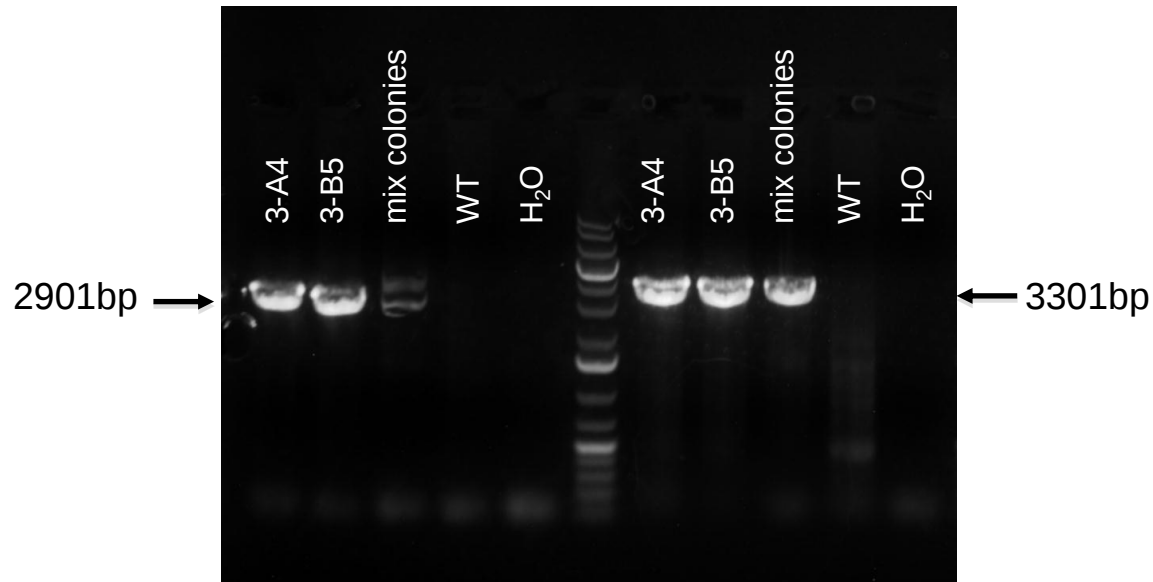
Conclusion

Genotyping results suggest that colonies 1-F6 and 1-C8 are positive *CL-CYH-008* KI cell lines. These colonies are ready for delivery and further analysis.

Genotyping-Junction PCR(the 2nd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

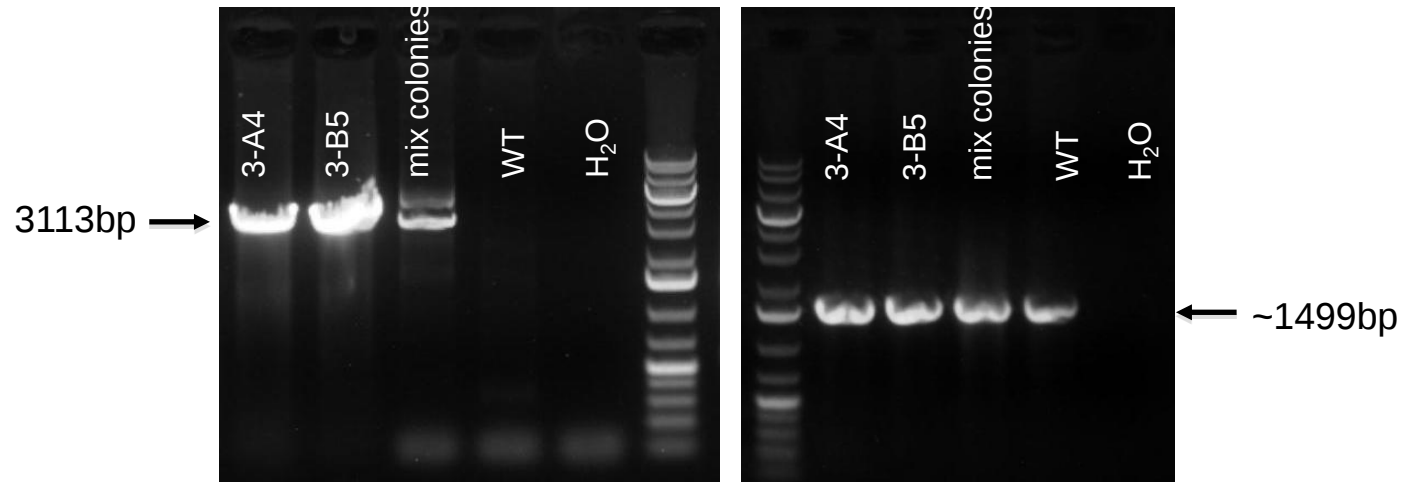
Primers: CL-CYH-008-R-GT-F/Puro-GT-F



Genotyping-Junction PCR(the 2nd time)

Primers: Pu Δ TK-SEQ-F1/CL-CYH-008-R-GT-R

Primers: CL-CYH-008-L-GT-F/ CL-CYH-008-FS-R



Colony screening by PCR and sequencing

Abbreviation:

HR: Homologous Recombination.

Δ : deletion.

WT: Wide type allele

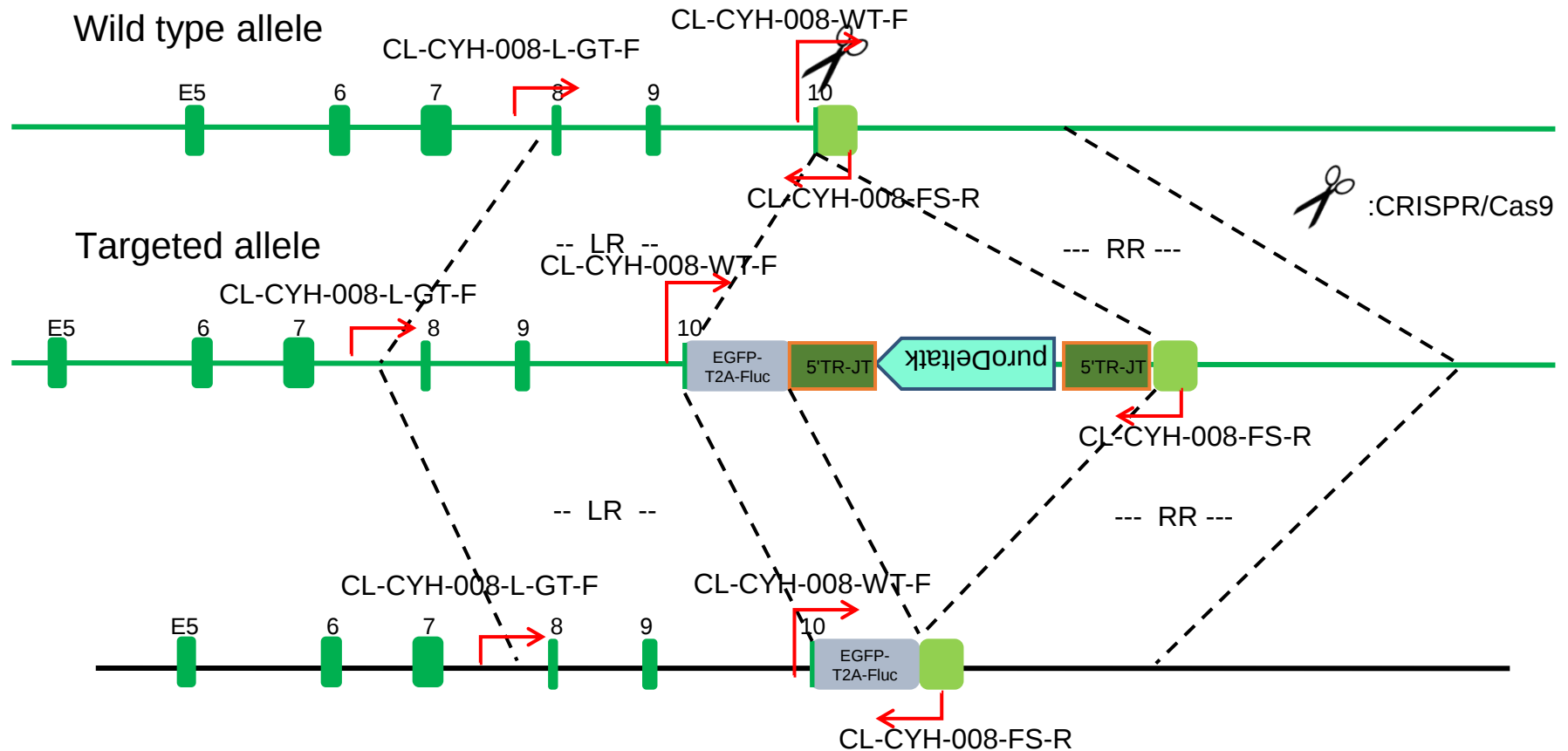
Colonies screening by PCR and sequencing(the 2nd time)

Colony	HR allele	non-HR allele	Genotype
3-A4	+	$\Delta 11\text{bp}$	KI, $\Delta 11$
3-B5	+	G $\rightarrow$ C	KI, WT(G $\rightarrow$ C)

Conclusion(the 2nd time)

Genotyping results suggest that colonies 3-A4 and 3-B5 are positive *CL-CYH-008* KI cell lines. These colonies are ready for delivery and further analysis.

III. Genotyping primer design-piggyBac transposase



III. Genotyping primer design-piggyBac transposase

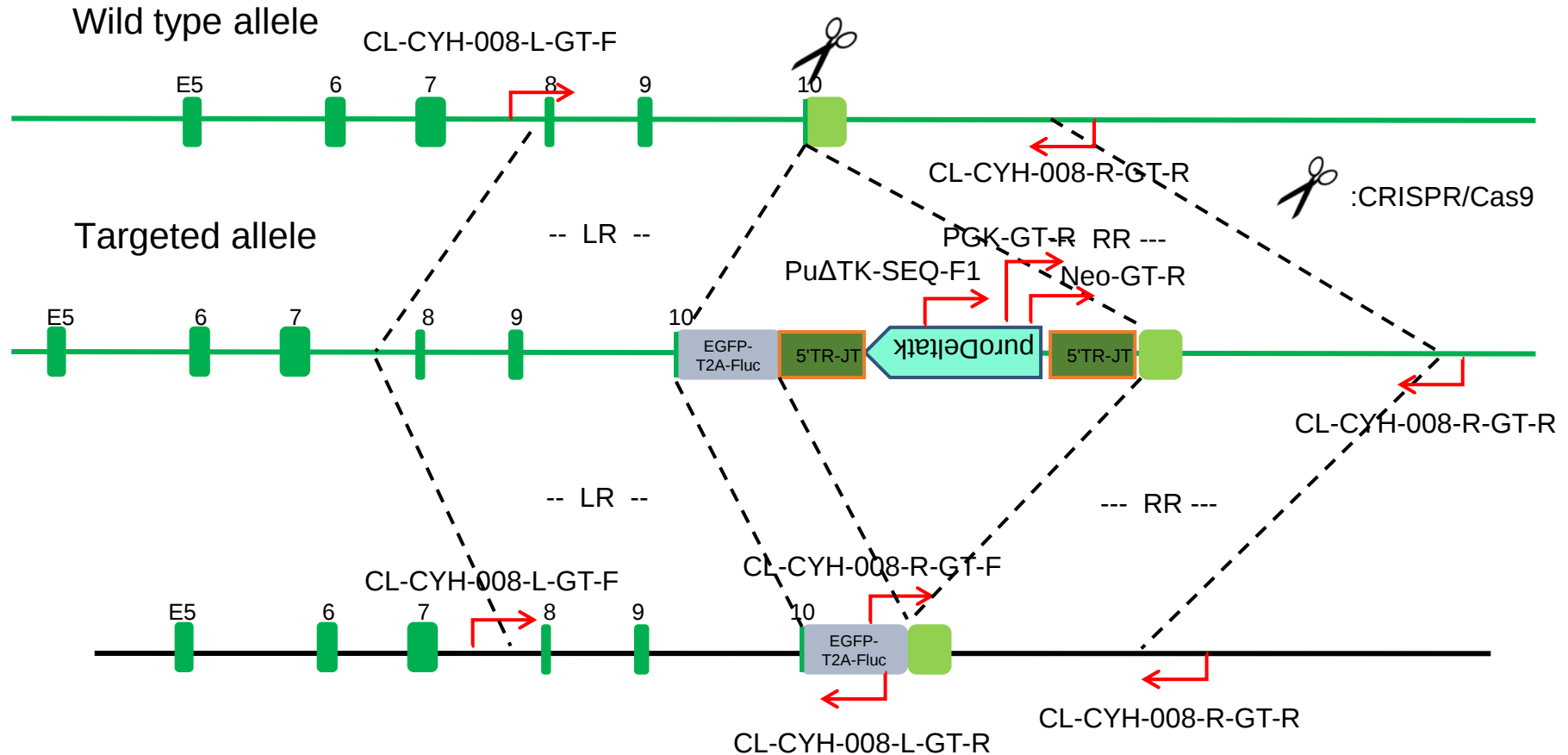
Primer	Sequence (5'-3')	Tm(°C)	Product size
CL-CYH-008-WT-F	GTCTTGTGTGTCTGCCCCTTCTAGT	62	Mut:2972bp WT:527bp Indel:~527bp
CL-CYH-008-FS-R	CCAAAGATTTATTGAAGCAGAACCAAGT	59	
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut:3944bp WT:1499bp Indel:~1499bp
CL-CYH-008-FS-R	CCAAAGATTTATTGAAGCAGAACCAAGT	59	

Enzyme: KOD-FX

Program: Touchdown PCR

94 °C	2 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

IV. Genotyping Junction PCR-piggyBac transposase



IV. Genotyping Junction PCR-piggyBac transposase

Primer	Sequence (5'-3')	Tm(°C)	Product size
CL-CYH-008-L-GT-F	TCCTGCTGCAAGTACTATCTCATCC	59	Mut: 2901bp
CL-CYH-008-L-GT-R	GTTGCTTAGGTCGTACTTGTCGATG	59	
CL-CYH-008-R-GT-F	ATGGATAGCAAGACCGACTACCAGG	61	Mut: 2680bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	60	

Enzyme: KOD-FX

Program: Touchdown PCR

94 °C	2 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

IV. Genotyping Junction PCR-piggyBac transposase

Primer	Sequence (5'-3')	Tm(°C)	Product size
Neo-GT-R	CAGAGGCCACTTGTGTAGCG	61	none(by piggyBac) Or 2062bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	59	
PuΔTK-SEQ-F1	AGTAGCGTGGGCATGGATCC	61	none(by piggyBac) Or 3113bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	59	
PGK-GT-R	AGAAAGCGAAGGAGCAAAGCTGCTA	63	none(by piggyBac) Or 2313bp
CL-CYH-008-R-GT-R	CGAAGTTTTCACTCCAGAACACACA	59	

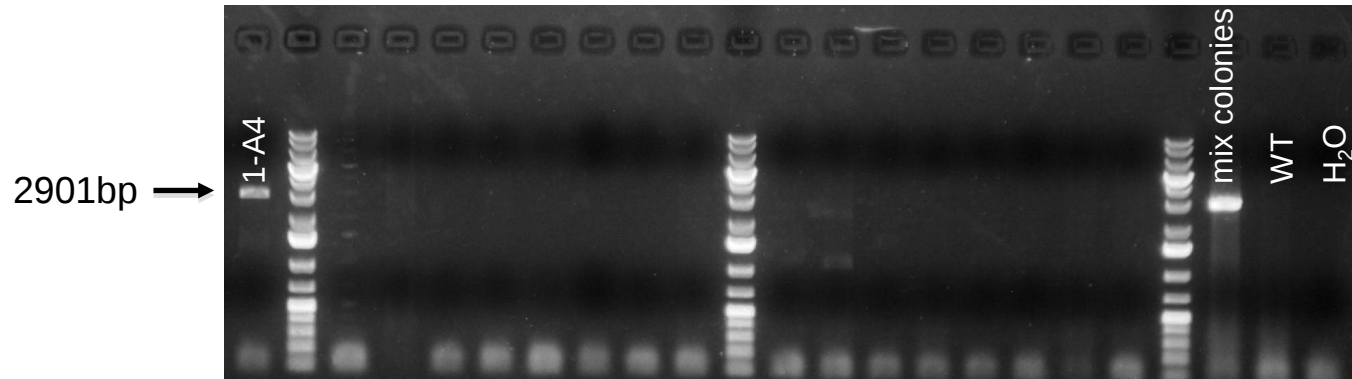
Enzyme: KOD-FX

Program: Touchdown PCR

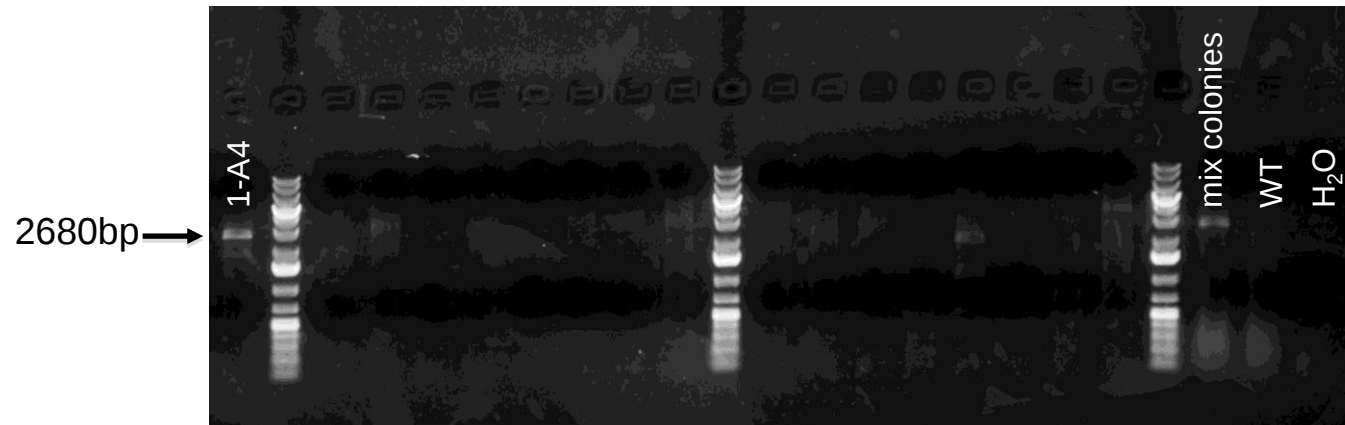
94 °C	2 min	} 15 cycles
98 °C	10 sec	
67 °C	30 sec (- 0.7°C/cycle)	
68 °C	1 kb / min	
98 °C	10 sec	} 25 cycles
57 °C	30 sec	
68 °C	1 kb / min	
68 °C	10 min	
4 °C	forever	

Genotyping-Junction PCR-piggyBac

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

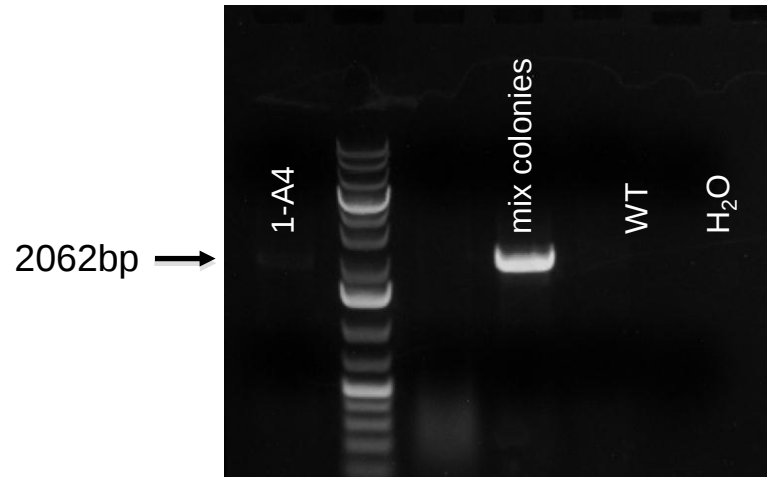


Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

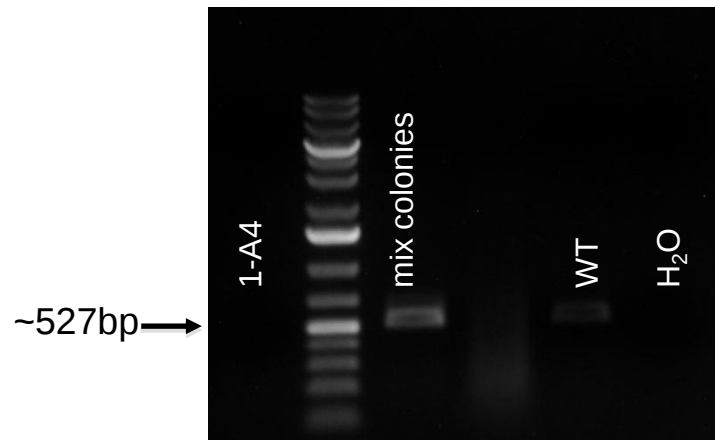


Genotyping-Junction PCR-piggyBac

Primers: Neo-GT-R/CL-CYH-008-R-GT-R



Primers: CL-CYH-008-WT-F/ CL-CYH-008-FS-R



Colony screening by PCR and sequencing

Abbreviation:

“no band”: No expected band was detected in PCR screening, but integration band was detected in the same reaction.

“none”: No WT/Indel sequence was detected in non-HR Allele PCR product.

HR: Homologous Recombination.

Colonies screening by PCR and sequencing

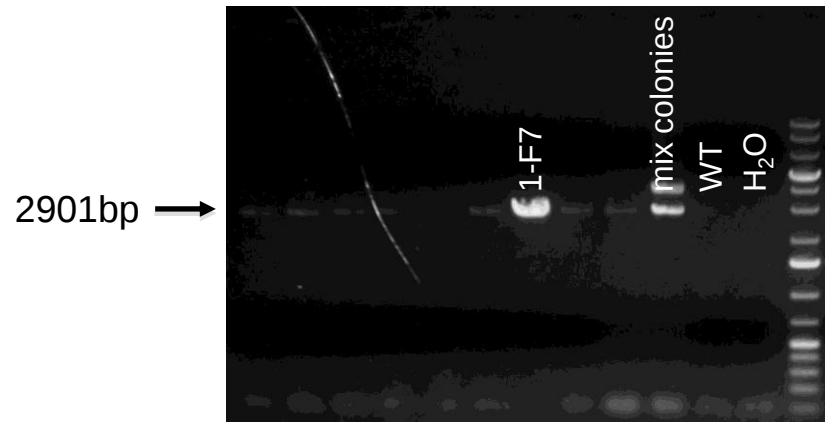
Colony	HR allele	non-HR allele	Genotype
1-A4	+	none	KI,KI

Conclusion

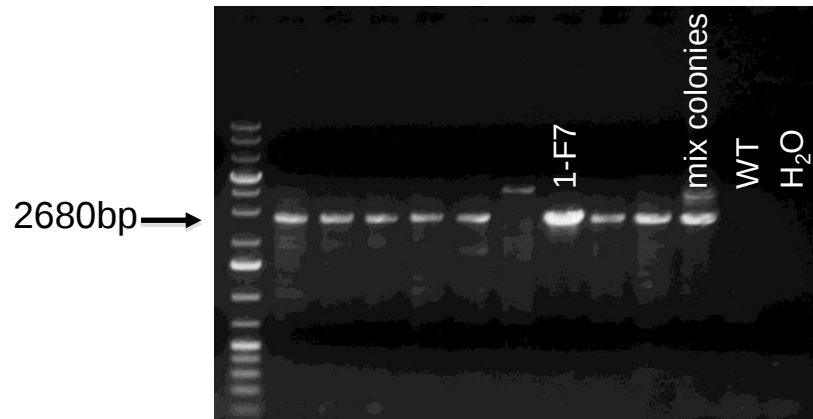
Genotyping results suggest that colony 1-A4 is a positive *CL-CYH-008* KI cell line. This colony is ready for delivery and further analysis.

Genotyping-Junction PCR-piggyBac(the 2nd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

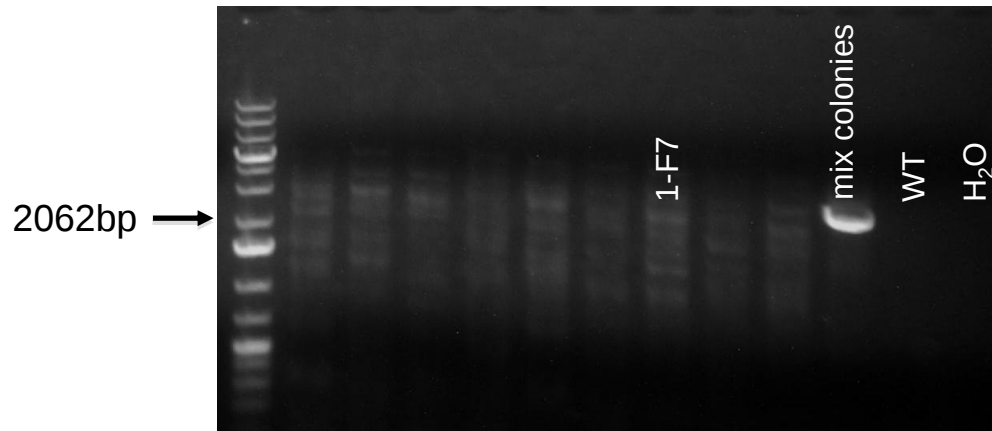


Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

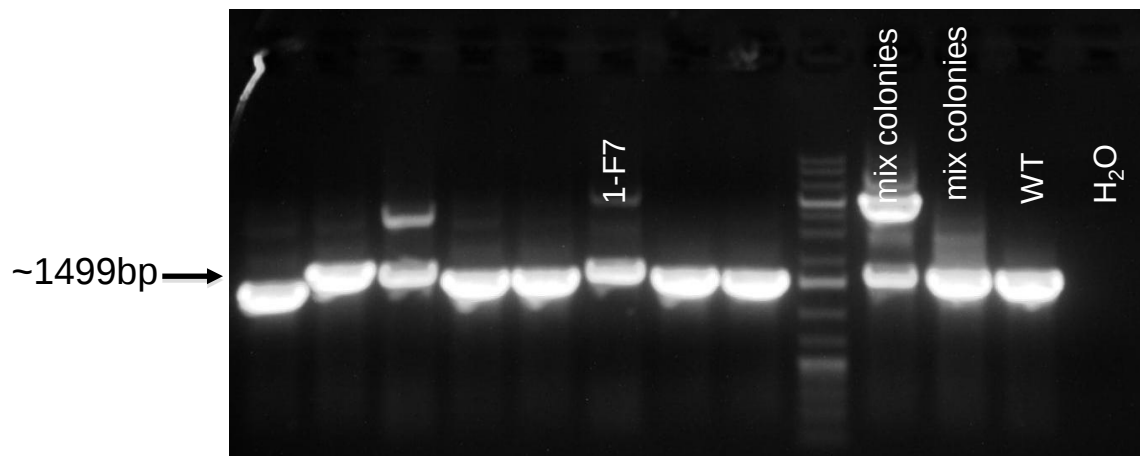


Genotyping-Junction PCR-piggyBac(the 2nd time)

Primers: Neo-GT-R/CL-CYH-008-R-GT-R



Primers: CL-CYH-008-L-GT-F/ CL-CYH-008-FS-R



Colony screening by PCR and sequencing

Abbreviation:

“no band”: No expected band was detected in PCR screening, but integration band was detected in the same reaction.

“none”: No WT/Indel sequence was detected in non-HR Allele PCR product.

FS: Frame shift.

HR: Homologous Recombination.

Colonies screening by PCR and sequencing(the 2nd time)

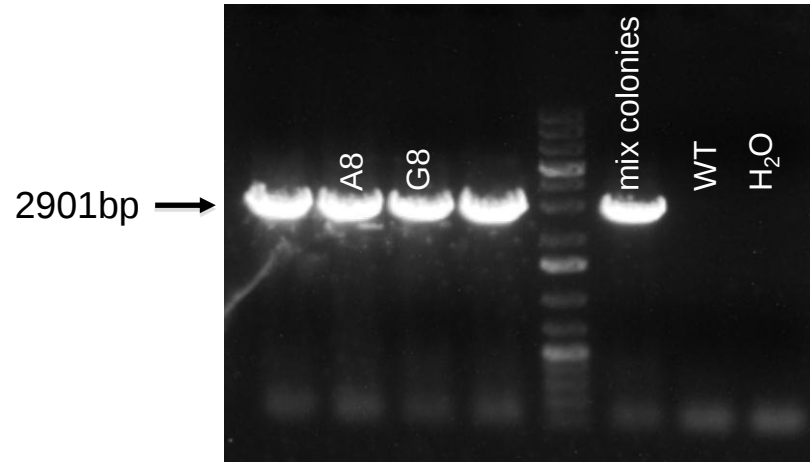
Colony	HR allele	non-HR allele	Genotype
1-F7	+	G→C	KI,WT(G→C)

Conclusion(the 2nd time)

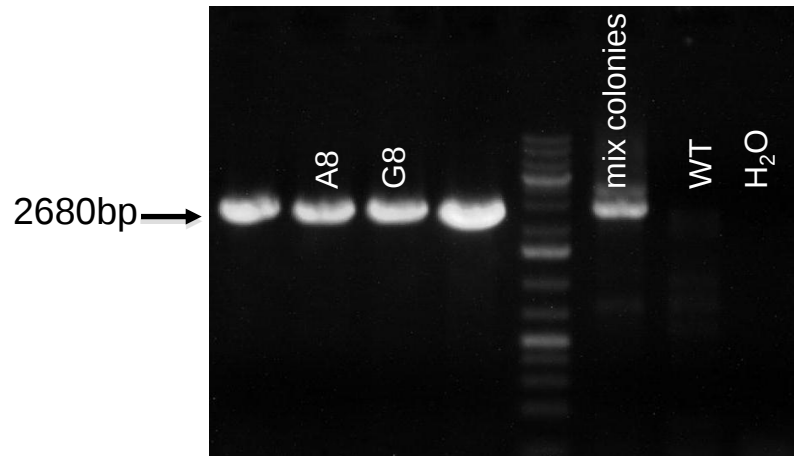
Genotyping results suggest that colony 1-F7 is a positive *CL-CYH-008* KI cell line. This colony is ready for delivery and further analysis.

Genotyping-Junction PCR-piggyBac(the 3rd time)

Primers: CL-CYH-008-L-GT-F/CL-CYH-008-L-GT-R

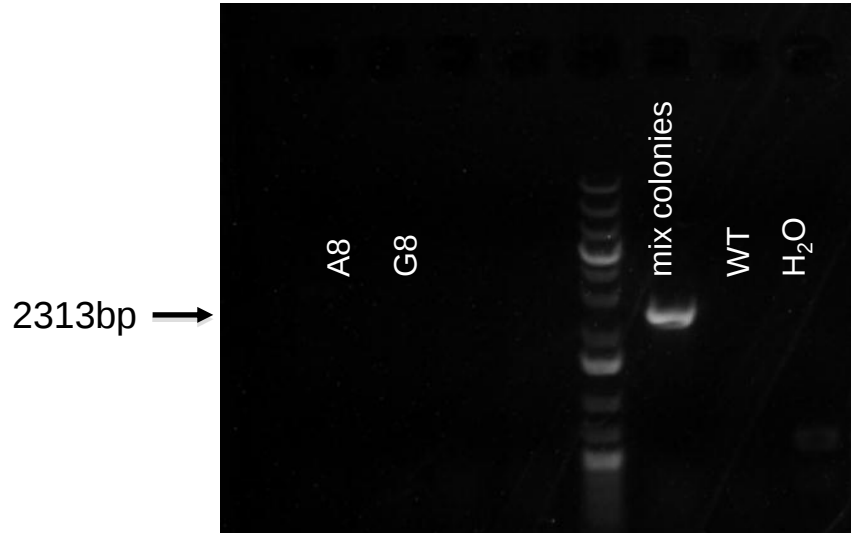


Primers: CL-CYH-008-R-GT-F/ CL-CYH-008-R-GT-R

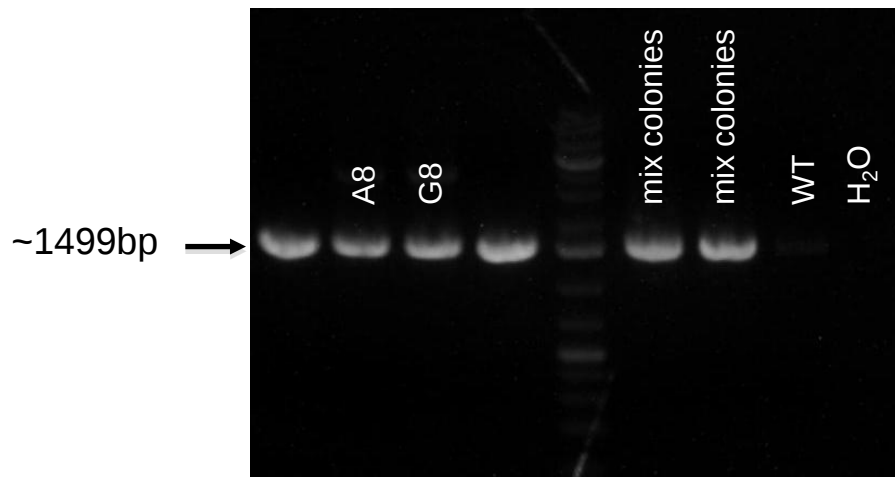


Genotyping-Junction PCR-piggyBac(the 3rd time)

Primers: PGK-GT-R/CL-CYH-008-R-GT-R



Primers: CL-CYH-008-L-GT-F/ CL-CYH-008-FS-R



Colony screening by PCR and sequencing

Abbreviation:

“no band”: No expected band was detected in PCR screening, but integration band was detected in the same reaction.

“none”: No WT/Indel sequence was detected in non-HR Allele PCR product.

HR: Homologous Recombination.

Colonies screening by PCR and sequencing(the 3rd time)

Colony	HR allele	non-HR allele	Genotype
A8	+	G→C	KI,WT(G→C)
G8	+	G→C	KI,WT(G→C)

Conclusion(the 3rd time)

Genotyping results suggest that colonies A8 and G8 are positive *CL-CYH-008* KI cell lines. These colonies are ready for delivery and further analysis.

This project has been completed.



Biocytogen was founded in 2008. Headquartered in Beijing, Biocytogen has two branches/facilities based in Haimen, Jiangsu China and Worcester MA, USA, also an administrative branch in Zhangjiang, Shanghai. Biocytogen is a global leader in providing high quality gene modified animal model generation and development, a top supplier of innovative human disease animal models in pharmaceutical research and a service provider of preclinical pharmacological/pharmacodynamics studies.

Biocytogen LLC
 info@biocytogen.com 1.508.421.4800
 One Innovation Drive, Worcester, MA 01605

